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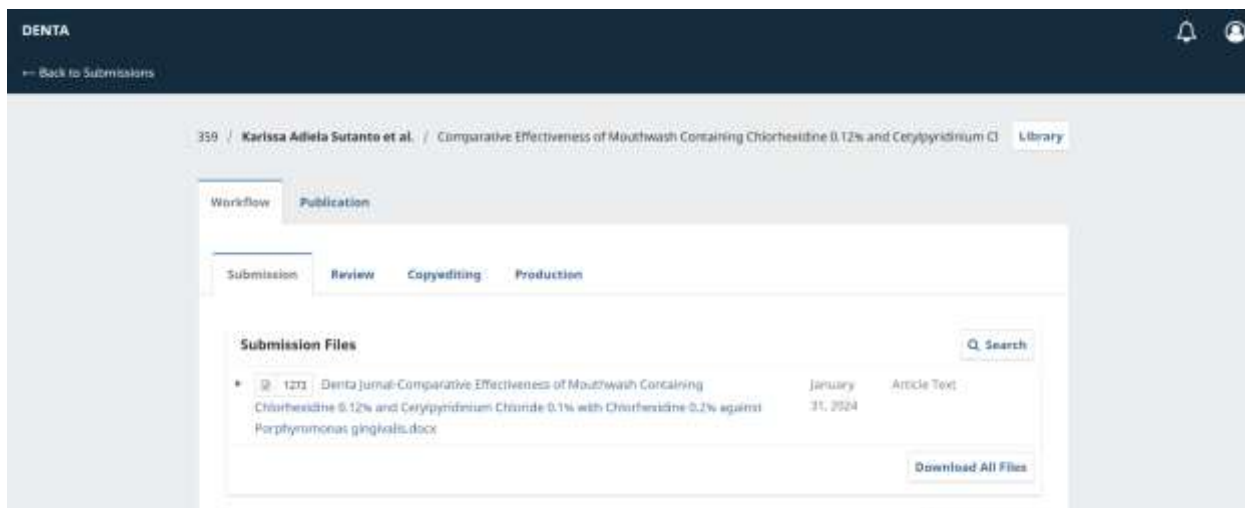
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Comparative Effectiveness of Mouthwash Containing Chlorhexidine 0.12% and Cetylpyridinium Chloride 0.1% with Chlorhexidine 0.2% against *Porphyromonas gingivalis*

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Abstract

Background : Periodontitis is an inflammatory disease that affects the supporting tissue of teeth that cause damage to the periodontal ligament and alveolar bone, generally caused by microorganisms. Chronic periodontitis is often caused by gram-negative anaerobic bacteria, especially *Porphyromonas gingivalis*. Mouthwash active ingredients that frequently used such as chlorhexidine (CHX) or cetylpyridinium chloride (CPC) have an antibacterial effect and prevent plaque formation. **Objective**: This research aims to compare the effectiveness of mouthwash containing CHX 0.12% and CPC 0.1% with CPC 0.2% on the bacterium *Porphyromonas gingivalis* ATCC 33277. **Methods** : This research is an experimental laboratory by giving treatment to *Porphyromonas gingivalis* with mouthwash CHX 0.12% and CPC 0.1%, CHX 0.2%, and distilled water as the negative control, then calculating the inhibition zone for each treatment and continued with One-Way ANOVA test. **Result** : The result showed the largest average of the inhibitory zone diameter was on CHX 0.2% which was 12,7 mm. Meanwhile, mouthwash containing CHX 0.12% and CPC 0.1% showed an average diameter of inhibitory zone resulted in

10,97 mm. **Conclusions** : In conclusion, CHX 0.2% has greater ability of inhibitory than mouthwash containing CHX 0.12% and CPC 0.1% against the growth of *Porphyromonas gingivalis*.

Keywords : antibacterial agents, cetylpyridinium chloride, chlorhexidine, mouthwash, *Porphyromonas gingivalis*

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Introduction

Oral health and hygiene are important things that need attention, because poor oral condition or inflammation can cause pain and discomfort.¹ Periodontal disease is one of the inflammatory condition that often found in oral cavity. The most common periodontal disease are gingivitis and periodontitis.² An inflammatory condition known as periodontitis affects the tissue that supports teeth and usually caused by certain bacteria, where there is damage to periodontal ligament and alveolar bone.³ The spread of inflammation from the epithelium to the connective tissue causes damage to the collagen fibers, followed by loss of attachment which is a sign of change from gingivitis to periodontitis. Periodontitis can be broadly classified into chronic and aggressive periodontitis.⁴ Chronic periodontitis is the most common type of periodontitis. The disease

progression of chronic periodontitis is slow to moderate and related to the plaque accumulation and calculus. The development of periodontitis disease may be caused by multifactorial, such as systemic, local or environmental factors that disrupt the interaction of normal host-bacteria.³ Although it can affect people of all ages, adults are more likely to be affected from chronic periodontitis. The level of local factors is correlated with the level of diseases progression. Certain bacteria cause chronic periodontitis to develop more slowly.⁴ Chronic periodontitis is often caused by gram-negative anaerobic bacteria, especially *Porphyromonas gingivalis*.⁵

Bacteria that often cause periodontal disease are *Porphyromonas gingivalis*, *Aggregibacter actinomycetemcomitans*, *Prevotella intermedia*, *Tannerella forsythia*,

Fusobacterium spp.⁶
Porphyromonas gingivalis is a gram-negative bacteria that often found in the subgingival area, and sometimes found in tongue and tonsils. The characteristics of *Porphyromonas gingivalis* are gram-negative, coccobacilli, non-motile, asaccharolytic and pleomorphic. *Porphyromonas gingivalis* grows anaerobically with dark pigmentation in media containing blood.^{6,7} *Porphyromonas gingivalis* has several virulence factors, such as gingipains, lipopolysaccharide (LPS), fimbriae, lectins (erythrocytes), capsules, collagenase, and proteases which release harmful and toxic metabolites and crucial in the early stages of periodontitis development.⁸

Periodontal disease can be prevented by maintaining oral hygiene, generally through plaque control by mechanical and chemical methods. Examples of mechanical methods are brushing teeth, using dental floss or an interdental brush. Examples of chemical methods are using toothpaste and mouthwash.⁹ The use of mouthwash can control supragingival plaque and gingivitis in individuals with a lack of motivation and skills to clean the oral cavity mechanically.¹⁰

Various kinds of mouthwash ingredients containing antimicrobials such as chlorhexidine and cetylpyridinium chloride, have shown efficacy in decreasing plaque and

preserving oral hygiene.¹¹ Chlorhexidine (CHX) is one of mouthwash with bisbiguanide ingredients that can kill microorganism by damaging their membrane cell, which damages the cytoplasm. Based on experimental studies, CHX is the gold standard for evaluating how effectively other mouthwashes work. Long-term use of CHX needs to be considered because it has the potential to cause staining on teeth and changes in taste.¹² Side effects that are often complained by the patients are stains on the teeth, mouth and buccal mucosa. There is also irritation of the oral mucosa, burning sensation and changes in taste perception.¹³ Side effects of CHX use are usually proportional to the duration of treatment.¹⁴

Chlorhexidine (CHX) is available in concentrations of 0.12% and 0.2% which affect plaque inhibition, the plaque inhibitory properties diminishing at lower concentrations.¹³ CHX 0.2% is bactericidal and CHX 0.12% is bacteriostatic. The decrease in CHX concentration is to reduce side effects while maintaining the effectiveness of the ingredient.¹⁵ CHX 0.2% is effective for preventing plaque and gingivitis.¹⁶

Cetylpyridinium chloride (CPC) is one of the mouthwash's active ingredients, that is made up of quaternary ammonium compounds, which are known to inhibit the growth

of bacteria. CPC can also be used as a treatment for halitosis. CPC with a concentration of 0.05%-0.1% effectively acts as an antimicrobial. A further approach for preventing periodontal disease is to use CPC as an antibacterial ingredient in mouthwash since it is considered to be safe, effective and has no serious adverse effects.⁸ CPC can cause extrinsic staining effects but only slightly compared to CHX mouthwash, because CPC is available in preparation alcohol-free, so the side effects that occur are less than CHX and more beneficial for all individuals.¹⁶

There is a combination of CHX and CPC mouthwash with the aim of eliminating the side effects of CHX and being more effective in inhibiting the growth of *Porphyromonas gingivalis* bacteria than using CHX mouth wash alone. This research aims to compare the effectiveness of CHX 0.12% and CPC 0.1% mouthwash with CHX 0.2% against *Porphyromonas gingivalis*.

Methods

This research used an experimental with post-test only control group design. This research was done at Microbiology Laboratory at Padjadjaran University in November-December 2023. *Porphyromonas gingivalis* bacteria on Mueller Hinton Agar (MHA) media were treated with a combination of

CHX 0.12% and CPC 0.1%, CHX 0.2%, and distilled water as negative control. The results observed were the inhibitory zone's diameter in millimeters. Data analysis was measured using the normality test, if the data is normally distributed, it will be tested using *One Way ANOVA* parametric test.

The sample for this research was *Porphyromonas gingivalis* ATCC 33277 acquired from Microbiology Laboratory of Padjadjaran University. To calculate the sample, the Federer formula is used, which $(t-1)(n-1) \geq 15$, where t is the amount of treatments, while n is the number of repetitions in each treatment, the result shows that $n \geq 8.5 \approx 9$. The number of repetitions for each treatment group is nine repetitions.

The tools used in this research were first sterilized with autoclave at 121°C for 15 minutes. In this research, using Mueller Hinton Agar (MHA) as the media, weighed 38 grams of MHA media, dissolved in 1L of distilled water until all the media was completely dissolved, sterilized using an autoclave at 121°C for 15 minutes. Preparation of microorganisms test by inoculating and culturing *Porphyromonas gingivalis* ATCC 33277 obtained from the Microbiology Laboratory, Padjadjaran University, Bandung, then incubated for 12-24 hours at 37°C.

Preparation of *Porphyromonas gingivalis* suspension was done by inoculating *Porphyromonas gingivalis* colonies that had been cultured on MHA media into bulyon solution, homogenized using a vortex mixer. The suspension's turbidity was adjusted to the standard solution McFarland 0.5 to obtain an inoculum with bacterial counts in the range of 1.5×10^8 CFU/mL.

The bacterial suspension was then put into the MHA media and spread on the surface of MHA using a sterile cotton swab, then treated with well diffusion. Well diffusion is done by making holes using a perforator in MHA media that has been inoculated with bacteria. Each hole was filled with treatment group, namely CHX 0.12% and CPC 0.1%, CHX 0.2%, and distilled water for the negative control. After that, the petri dish was incubated in an incubator at

37°C for 24 hours. The same procedure is repeated nine times, then observe and calculate the inhibitory zone's diameter that formed around the hole using caliper.

Antibacterial activity can be categorized from the diameter of inhibition zone, which is weak with the diameter <5 mm, moderate 5-10 mm, strong 10-20 mm, and very strong with an inhibition zone diameter >20 mm.¹⁷

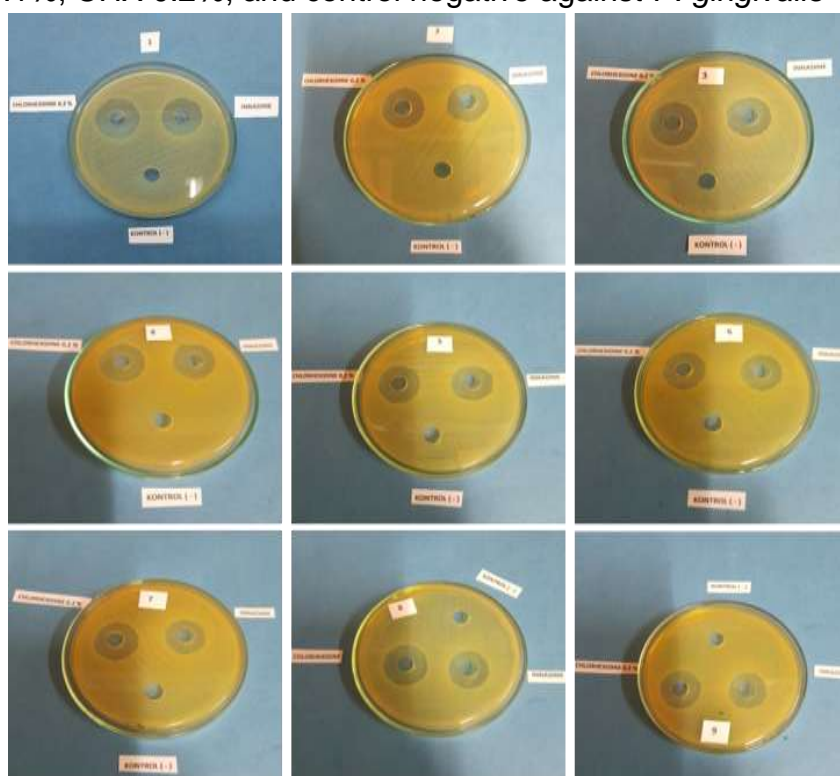
Results

The results of measuring the diameter of the inhibitory zone for mouthwash containing CHX 0.12% and CPC 0.1%, CHX 0.2%, and distilled water against *Porphyromonas gingivalis* can be seen in table 1.

Table 1. Results of Measurement of Inhibitory Zone Diameter

Inhibitory Zone Diameter	Treatment		
	CHX 0.12% dan CPC 0.1% (mm)	CHX 0.2% (mm)	Negative control
1	11.57	14.55	0.00
2	10.80	12.1	0.00
3	11.05	13.42	0.00
4	11.35	12.82	0.00
5	11.15	12.65	0.00
6	11.52	12.02	0.00
7	9.7	12.05	0.00
8	10.85	13.6	0.00
9	10.72	11.05	0.00
Mean (mm)	10.97	12.7	0

Figure 1. The results of observation of the inhibitory zone diameter of CHX 0.12% and CPC 0.1%, CHX 0.2%, and control negative against *P. gingivalis*



Based on research, the largest diameter of the inhibition zone was in the CHX 0.2% mouthwash treatment, which was 12.7 mm. The next diameter of inhibition zone is in the mouthwash treatment containing CHX 0.12% and CPC 0.1%, which was 10.97 mm. While in the negative control treatment, no inhibition zone was developed or the inhibition zone was zero. Based on the classification of inhibition, the result of measurement

of the inhibitory zone diameter for CHX 0.12% and CPC 0.1% and CHX 0.2% included in the strong category (Table 2).

Table 2. Classification of inhibition zones of CHX 0.12% and CPC 0.1% and CHX 0.2% against *P. gingivalis*

Treatment	Diameter mean (mm)	Classification of Inhibition Zone
CHX 0.12% dan CPC 0.1%	10.97	Strong
CHX 0.2%	12.7	Strong

The normality test was done using *Shapiro-Wilk* because the samples were less than 50. The normality test results for CHX 0.12% and CPC 0.1% showed a significance value of 0.138 and for CHX 0.2% showed significance value of 0.905. The data results show a $p\text{-value} > 0.05$, it can be conclude that the data is normally distributed, then it can be continued with analysis statistic using *One Way ANOVA* test. After normality test, continued with homogeneity test using the *Levene* test to find out whether the data is homogeneous or not. The homogeneity test results show a significance value of 0.002 ($p\text{-value} < 0.05$) so it can be assumed that the data is not homogeneous.

The results of *One Way ANOVA* test showed a significance value of 0.000 where $p\text{-value} < 0.05$, means there were significant differences in the three treatments given. The non-homogeneous data was continued with *Post Hoc* test using the *T-test* to determine which treatment was the most significant. *Post Hoc* test results showed $p\text{-value} < 0.05$. It can be concluded that CHX

0.12% and CPC 0.12%, CHX 0.2%, and distilled water were significantly difference in inhibiting *Porphyromonas gingivalis*.

Discussion

This research used well diffusion method to see the difference in effectiveness between mouthwash containing CHX 0.12% and CPC 0.1% with CHX 0.2%. The larger the clear zone formed around the hole, the higher the inhibition. From the results of the inhibition zone measurements, the largest average of inhibitory zone's diameter is on mouthwash containing CHX 0.2%. This shows that CHX 0.2% is the most effective mouthwash in inhibiting *Porphyromonas gingivalis* compared to mouthwash containing CHX 0.12% and CPC 0.1%.

Based on the research by Betadion R, *et al.*, chlorhexidine has the strongest antibacterial effect against *Porphyromonas gingivalis*, so CHX is used as the gold standard and often as a positive control for antibacterial examination of other materials.¹⁸ CHX is considered as gold standard for antimicrobial

mouthwash due to its proven long-term effectiveness. However, due to the side effects of CHX such as staining/discoloration of teeth and oral mucosa, unpleasant taste, and alcohol content, certain individuals cannot use CHX and it can only be used in the short term.

The mechanism of action of CHX as an antibacterial is described with the damage to bacterial cell membrane and leakage of cytoplasmic components. The positive charge (cations) in CHX attach to the negative charge of bacterial molecules (anions). This will cause changes in the cell membrane which will disrupt the permeability of bacterial cell wall, then release the intercellular fluid or leakage of components and resulting in cell death. The higher the concentration of CHX, the greater the damage to the bacterial cell membrane that occurs. At low and high concentrations, CHX is bacteriostatic (inhibits bacterial growth) and bactericidal (kills bacteria).¹⁸

CPC plays a role in antimicrobial activity on supragingival plaque bacteria.¹⁹ Several studies examining the combination efficacy of CPC and CHX and showed a decrease in plaque levels and numbers of bacteria, as well as bleeding on probing (BOP) scores. The mouthwash combination of CPC and CHX may be effective for long-term

use, due to the lower concentrations of CHX. It is hoped that this combination of mouthwash can maintain its effectiveness by minimizing the side effects that will occur.

The mechanism of action of CPC as an antibacterial is by increasing interaction with bacterial cells and causing damage to the cell membrane resulting in leakage of cytoplasmic components, metabolic disorders and eventually cause death of bacteria. At low concentrations, CPC affects cells by disrupting osmoregulation and homeostasis. At high concentrations, CPC causes membrane damage and leakage of cytoplasmic components. CPC can also inhibit the synthesis of glucans that it can inhibit the formation of biofilms.²⁰

Based on research by Stela Lima F, *et al.*, there are limitations to the use of CPC, where the *Porphyromonas gingivalis* only decreases with the use of CHX. Levels of periodontopathogens in CPC-treated biofilms were statistically similar to untreated biofilms. CPC only reduced 50% levels of *Porphyromonas gingivalis*.²¹ In addition, the concentration of CHX in CHX 0.2% was higher compared to the combination of CHX 0.12% and CPC 0.1%. These things allow the use of CHX 0.2% to be better than CHX 0.12% and CPC 0.1% against *Porphyromonas gingivalis*.

There are various other factors that affect the results of the differences in the inhibition zones formed. Factors that can affect the inhibition zone include the sensitivity of the organism, incubation temperature, incubation time, turbidity of the bacterial suspension, and thickness of the agar medium. The incubation temperature must be done at 37°C and for 24 hours, 1. temperatures less than 37°C can cause a larger inhibition zone diameter. In bacterial suspension 2. turbidity, the diameter of inhibition zone will be larger if the suspension is not more turbid than the turbidity of standard McFarland 0.5, and will be smaller if the suspension is more turbid. Less thickness of agar media can cause the diffusion process to be faster and vice versa.²²

Conclusion

Based on the results of the research, it can be concluded that CHX 0.2% mouthwash has greater inhibitory 4. zone than CHX 0.12% and CPC 0.1% mouthwash against 5. *Porphyromonas gingivalis*. There is a significant difference between the effectiveness of mouthwash containing CHX 0.12% and CPC 0.1% with CHX 0.2% mouthwash in inhibiting the growth of 6. *Porphyromonas gingivalis*.

Acknowledgement

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Conflicts of Interest

The authors declare no conflict of interest.

References

- Lang NP, Bartold PM. Periodontal Health. J Periodontol. 2018; 89(1):9–16. DOI: 10.1002/JPER.16-0517.
- Achmad H, Singgih MF, Huldani, Ramdhani AF, Ramadhany YF. Inhibitory Power Test of White Rice Bran Extract (*Oryza sativa L.*) with the Solution of Ethanol and Aquades on *Porphyromonas gingivalis* (In Vitro) Bacteria. Systematic Reviews in Pharmacy. 2020; 11(6):858–63. DOI: 10.31838/srp.2020.6.123.
- Carranza FA, Newman MG, Takei HH, Klokkevold PR. Carranza's Clinical Periodontology. 13th ed. Philadelphia: Elsevier; 2019.
- Mehrotra N, Singh S. Periodontitis. StatPearls Publishing. 2023.
- Rafiei M, Kiani F, Sayehmiri F, Sayehmiri K, Sheikhi A, Azodi MZ. Study of *Porphyromonas gingivalis* in periodontal diseases: A systematic review and meta-analysis. Med J Islam Repub Iran. 2017; 31(62):2–5. DOI: 10.18869/mjiri.31.62.
- Marsh PD, Lewis MA, Rogers H, Williams DW, Wilson M. Marsh and Martin's Oral Microbiology. 6th ed. Marsh and Martin's Oral Microbiology. UK: Elsevier; 2016.

7. Samaranayake L. Essential Microbiology for Dentistry. 5th ed. Essential Microbiology for Dentistry. Churchill Livingstone Elsevier; 2018.
8. Setiawatie EM, Valentina R, Meiliana RS. Effectiveness of Cetylpyridinium Chloride in Reducing the Growth of Bacteria that Cause Periodontal Disease. 2023; 11(2):115–20. DOI: 10.35790/eg.v11i.
9. Toar AI, Posangi J, Wowor V. Daya Hambat Obat Kumur Cetylpyridinium Chloride dan Obat Kumur Daun Sirih Terhadap Pertumbuhan *Streptococcus mutans*. Jurnal Biomedik. 2013; 5(1):163–8. DOI: 10.35790/jbm.5.1.2013.2639.
10. Farah CS, McIntosh L, McCullough MJ. Mouthwashes. Aust Prescr. 2013; 32(6):162–4. DOI: 10.18773/austprescr.2009.080.
11. Yarahmadi N, Hashemian F, Doust HH. Clinical Effects of Chlorhexidine 0.2% and Cetylpyridinium 0.05% Combination in Comparison with Chlorhexidine, Cetylpyridinium and Persica in Reducing Oral Bacteria in Healthy Individuals. J Pharm Care. 2020; 8(3):116–22. DOI: 10.18502/jpc.v8i3.4545.
12. Adkins KL. Effectiveness of cetylpyridinium chloride, chlorhexidine gluconate, chlorine dioxide, and essential oils against *F. nucleatum*, *P. gingivalis*, *S. mutans* and *S. sobrinus* - a biofilm approach. 2013; 5(1):15–23.
13. Hassan Najafi M, Taheri M, Reza Mokhtari M, Forouzanfar A, Farazi F, Mirzaee M, et al. Comparative study of 0.2% and 0.12% digluconate chlorhexidine mouth rinses on the level of dental staining and gingival indices. Dent Res J (Isfahan). 2013; 9(3):305–8.
14. Tartaglia GM, Tadakamadla SK, Connelly ST, Sforza C, Martín C. Adverse events associated with home use of mouthrinses: a systematic review. Ther Adv Drug Saf. 2019; 10(1):1–16. DOI: 10.1177/2042098619854881.
15. Dutt P, Kr Rathore P, Khurana D. Chlorhexidine - An antiseptic in periodontics. IOSR Journal of Dental and Medical Sciences. 2014; 13(9):85–8. DOI: 10.4103/jpbs.JPBS_162_20.
16. Sari D Novita, Cholil, Sukmana B Indra. Perbandingan Efektifitas Obat Kumur Bebas Alkohol Yang Mengandung Cetylpyridinium Chloride Dengan Chlorhexidine Terhadap Penurunan Plak. Dentino. 2014; 2(2):179–83.
17. Tille PM. Bailey & Scott's Diagnostic Microbiology. 13th ed. St. Louis, Missouri : Elsevier; 2014.
18. Sinaredi BR, Pradopo S, Wibowo TB. Daya antibakteri obat kumur chlorhexidine, povidone iodine, fluoride suplementasi zinc terhadap *Streptococcus mutans* dan *Porphyromonas gingivalis*. Dent J. 2014; 47(4):211–4. DOI: 10.20473/j.djmk.v47.i4.p211-214.
19. Sreenivasan PK, Haraszthy VI, Zambon JJ. Antimicrobial efficacy of 0.05% cetylpyridinium chloride mouthrinses. Lett Appl Microbiol.

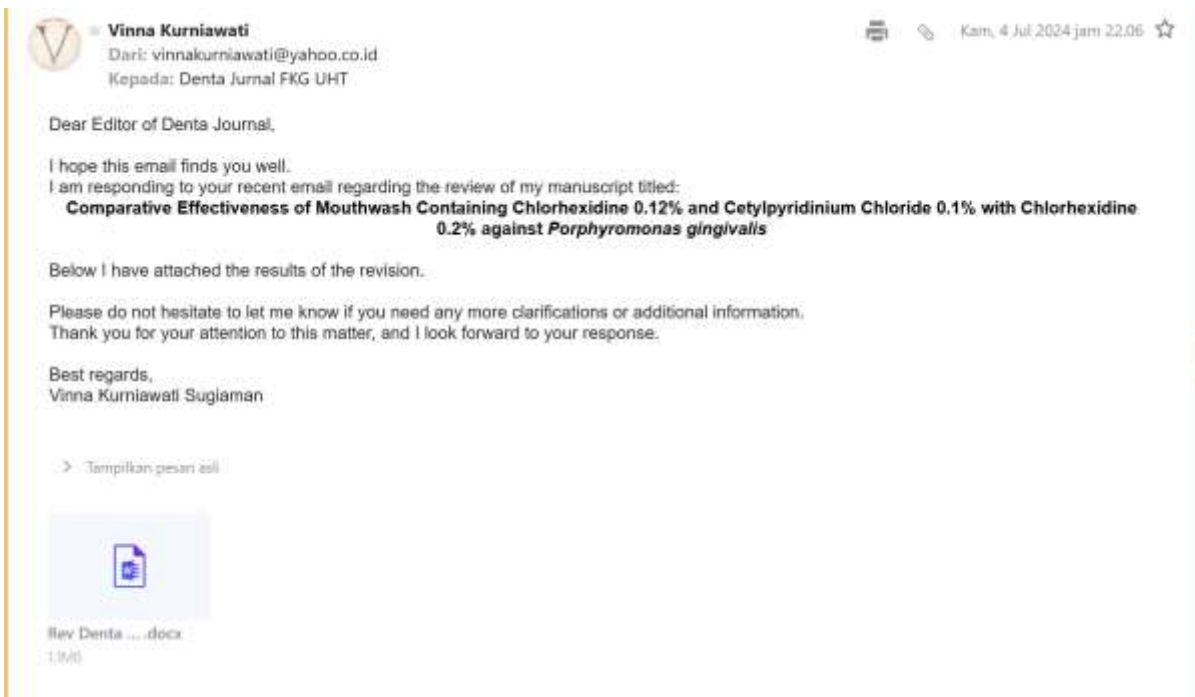
2013; 56(1):14–20. DOI:
10.1111/lam.12008.

20. Mao X, Auer DL, Buchalla W, Hiller KA, Maisch T, Hellwig E, et al. Cetylpyridinium Chloride: Mechanism of Action, Antimicrobial Efficacy in Biofilms, and Potential Risks of Resistance. *Antimicrob Agents Chemother.* 2020; 64(8):2–4. DOI: 10.1128/AAC.00576-20.
21. De Miranda SLF, Damaceno JT, Faveri M, Figueiredo LC, Soares GMS, Bueno-Silva B. In Vitro Antimicrobial Effect of Cetylpyridinium Chloride on Complex Multispecies Subgingival Biofilm. *Braz Dent J.* 2020; 31(2):103–8. DOI: 10.1590/0103-6440202002630.
22. Zeniusa P, Ricky Ramadhian M, Hamidi Nasution S, Karima N. Uji Daya Hambat Ekstrak Etanol Teh Hijau Terhadap *Escherichia coli* Secara In Vitro. *Majority.* 2019; 8(2):136–43.

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ABSTRACT

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INTRODUCTION

Oral health and hygiene are important things that need attention, because poor oral condition or inflammation can cause pain and discomfort.¹ Periodontal disease is one of the inflammatory condition that often found in oral cavity. The most common periodontal disease are gingivitis and periodontitis.² An inflammatory condition known as periodontitis affects the tissue that supports teeth and usually caused by certain bacteria, where there is damage to periodontal ligament and alveolar bone.³ The spread of inflammation from the epithelium to the connective tissue causes damage to



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Various kinds of mouthwash ingredients containing antimicrobials such as chlorhexidine and cetylpyridinium chloride, have shown efficacy in decreasing plaque and preserving oral hygiene.¹¹ Chlorhexidine (CHX) is one of mouthwash with bisbiguanide ingredients that can kill microorganism by damaging their membrane cell, which damages the cytoplasm. Based on experimental studies, CHX is the gold standard for evaluating how effectively other mouthwashes work. Long-term use of CHX needs to be considered because it has the potential to cause staining on teeth and changes in taste.¹² Side effects that are often complained by the patients are stains on the teeth, mouth and buccal mucosa. There is also irritation of the oral mucosa, burning sensation and changes in taste perception.¹³ Side effects of CHX use are usually proportional to the duration of treatment.¹⁴

Chlorhexidine (CHX) is available in concentrations of 0.12% and 0.2% which affect plaque inhibition, the plaque inhibitory properties diminishing at lower concentrations.¹³ CHX 0.2% is bactericidal and CHX 0.12% is bacteriostatic. The decrease in CHX concentration is to reduce side effects while maintaining the effectiveness of the ingredient.¹⁵ CHX 0.2% is effective for preventing plaque and gingivitis.¹⁶

Cetylpyridinium chloride (CPC) is one of the mouthwash's active ingredients, that is made up of quaternary ammonium compounds, which are known to inhibit the growth of bacteria. CPC can also be used as a treatment for halitosis. CPC with a concentration of 0.05%-0.1% effectively acts as an antimicrobial. A further approach for preventing periodontal disease is to use CPC as an antibacterial ingredient in mouthwash since it is considered to be safe, effective and has no serious adverse effects.⁸ CPC can cause extrinsic staining effects but only slightly compared to CHX mouthwash, because CPC is available in preparation alcohol-free, so the side effects that occur are less than CHX and more beneficial for all individuals.¹⁶

There is a combination of CHX and CPC mouthwash to eliminate the side effects of CHX and be more effective in inhibiting the growth of *Porphyromonas gingivalis* bacteria than using CHX



mouth wash alone. This research aims to compare the effectiveness of CHX 0.12% and CPC 0.1% mouthwash with CHX 0.2% against *Porphyromonas gingivalis*.

MATERIALS AND METHODS

This research used an experimental with post-test only control group design. This research was done at Microbiology Laboratory at Padjadjaran University in November-December 2023. *Porphyromonas gingivalis* bacteria on Mueller Hinton Agar (MHA) media were treated with a combination of CHX 0.12% and CPC 0.1%, CHX 0.2%, and distilled water as negative control. The results observed were the inhibitory zone's diameter in millimeters. Data analysis was measured using the normality test, if the data is normally distributed, it will be tested using *One Way ANOVA* parametric test.

The sample for this research was *Porphyromonas gingivalis* ATCC 33277 acquired from Microbiology Laboratory of Padjadjaran University. To calculate the sample, the Federer formula is used, which $(t-1)(n-1) \geq 15$, where t is the amount of treatments, while n is the number of repetitions in each treatment, the result show that $n \geq 8.5 \approx 9$. The number of repetitions for each treatment group is nine repetitions.

The tools used in this research was first sterilized with autoclave at 121°C for 15 minutes. In this research, using Mueller Hinton Agar (MHA) as the media, weighed 38 grams of MHA media, dissolved in 1L of distilled water until all the media was completely dissolved, sterilized using an autoclave at 121°C for 15 minutes. Preparation of microorganisms test by inoculating and culturing *Porphyromonas gingivalis* ATCC 33277 obtained from the Microbiology Laboratory, Padjadjaran University, Bandung, then incubated for 12-24 hours at 37°C.

Preparation of *Porphyromonas gingivalis* suspension was done by inoculating *Porphyromonas gingivalis* colonies that had been cultured on MHA media into bulyon solution, homogenized using a vortex mixer. The suspension's turbidity was adjusted to the standard solution McFarland 0.5 to obtain an inoculum with bacterial counts in the range of 1.5×10^8 CFU/mL.

The bacterial suspension was then put into the MHA media and spread on the surface of MHA using a sterile cotton swab, then treated with well diffusion. Well diffusion is done by making holes using a perforator in MHA media that has been inoculated with bacteria. Each hole was filled with treatment group, namely CHX 0.12% and CPC 0.1%, CHX 0.2%, and distilled water for the negative control. After that, the petri dish was incubated in an incubator at 37°C for 24 hours. The same procedure is repeated nine times, then observe and calculate the inhibitory zone's diameter that formed around the hole using caliper.

Antibacterial activity can be categorized from the diameter of inhibition zone, which is weak with the diameter <5 mm, moderate 5-10 mm, strong 10-20 mm, and very strong with an inhibition zone diameter >20 mm.¹⁷

RESULT

The results of measuring the diameter of the inhibitory zone for mouthwash containing CHX 0.12% and CPC 0.1%, CHX 0.2%, and distilled water against *Porphyromonas gingivalis* can be seen in table 1.



Table 1. Results of Measurement of Inhibitory Zone Diameter

Inhibitory Zone Diameter	Treatment		
	CHX 0.12% dan CPC 0.1% (mm)	CHX 0.2% (mm)	Negative control
1	11.57	14.55	0.00
2	10.80	12.1	0.00
3	11.05	13.42	0.00
4	11.35	12.82	0.00
5	11.15	12.65	0.00
6	11.52	12.02	0.00
7	9.7	12.05	0.00
8	10.85	13.6	0.00
9	10.72	11.05	0.00
Mean (mm)	10.97	12.7	0

Figure 1. The results of observation of the inhibitory zone diameter of CHX 0.12% and CPC 0.1%, CHX 0.2%, and control negative against *P. gingivalis*



Based on research, the largest diameter of the inhibition zone was in the CHX 0.2% mouthwash treatment, which was 12.7 mm. The next diameter of inhibition zone is in the mouthwash treatment containing CHX 0.12% and CPC 0.1%, which was 10.97 mm. While in the negative control



treatment, no inhibition zone was developed or the inhibition zone was zero. Based on the classification of inhibition, the result of measurement of the inhibitory zone diameter for CHX 0.12% and CPC 0.1% and CHX 0.2% included in the strong category (Table 2).

Table 2. Classification of inhibition zones of CHX 0.12% and CPC 0.1% and CHX 0.2% against *P. gingivalis*

Treatment	Diameter mean (mm)	Classification of Inhibition Zone
CHX 0.12% dan CPC 0.1%	10.97	Strong
CHX 0.2%	12.7	Strong

The normality test was done using *Shapiro-Wilk* because the samples were less than 50. The normality test results for CHX 0.12% and CPC 0.1% showed a significance value of 0.138 and for CHX 0.2% showed significance value of 0.905. The data results show a $p\text{-value} > 0.05$, it can be concluded that the data is normally distributed, then it can be continued with analysis statistic using *One Way ANOVA* test. After normality test, continued with homogeneity test using the *Levene* test to find out whether the data is homogeneous or not. The homogeneity test results show a significance value of 0.002 ($p\text{-value} < 0.05$) so it can be assumed that the data is not homogeneous. The results of *One Way ANOVA* test showed a significance value of 0.000 where $p\text{-value} < 0.05$, means there were significant differences in the three treatments given. The non-homogeneous data was continued with *Post Hoc* test using the *T-test* to determine which treatment was the most significant. *Post Hoc* test results showed $p\text{-value} < 0.05$. It can be concluded that CHX 0.12% and CPC 0.12%, CHX 0.2%, and distilled water were significantly difference in inhibiting *Porphyromonas gingivalis*.

DISCUSSION

This research used well diffusion method to see the difference in effectiveness between mouthwash containing CHX 0.12% and CPC 0.1% with CHX 0.2%. The larger the clear zone formed around the hole, the higher the inhibition. From the results of the inhibition zone measurements, the largest average of inhibitory zone's diameter is on mouthwash containing CHX 0.2%. This shows that CHX 0.2% is the most effective mouthwash in inhibiting *Porphyromonas gingivalis* compared to mouthwash containing CHX 0.12% and CPC 0.1%.

Based on the research by Betadion R, *et al.*, chlorhexidine has the strongest antibacterial effect against *Porphyromonas gingivalis*, so CHX is used as the gold standard and often as a positive control for antibacterial examination of other materials.¹⁸ CHX is considered as gold standard for antimicrobial mouthwash due to its proven on long-term of effectiveness. However, due to the side effects of CHX such as staining/discoloration of teeth and oral mucosa, unpleasant taste, and alcohol content, certain individuals cannot use CHX and its can only be used in the short term.

The mechanism action of CHX as an antibacterial is described with the damage to bacterial cell membrane and leakage of cytoplasmic components. The positive charge (cations) in CHX attach the negative charge of bacterial molecules (anions). This will cause changes in the cell membrane which will disrupt the permeability of bacterial cell wall, then release the intercellular fluid or leakage of components resulting in cell death. The higher the concentration of CHX, the greater the damage to the bacterial cell membrane that occurs. At low and high concentrations, CHX is bacteriostatic (inhibits bacterial growth) and bactericidal (kills bacteria).¹⁸

CPC plays role in antimicrobial activity on supragingival plaque bacteria.¹⁹ Several studies examining the combination efficacy of CPC and CHX and showed a decrease result in plaque levels and numbers of bacteria, as well as bleeding on probing (BOP) scores. The mouthwash combination



of CPC and CHX may be effective for long-term use, due to the lower concentrations of CHX. It is hoped that this combination of mouthwash can maintain its effectiveness by minimizing the side effects that will occur.

The mechanism action of CPC as an antibacterial by increasing interaction with bacterial cells and causing damage to the cell membrane resulting in leakage of cytoplasmic components, metabolic disorders and eventually cause death cell of bacteria. At low concentrations, CPC affects cells by disrupting osmoregulation and homeostasis. At high concentrations, CPC cause membrane damage and leakage of cytoplasmic components. CPC can also inhibit the synthesis of glucans that it can inhibit the formation of biofilms.²⁰

Based on research by Stela Lima F, *et al.*, there are limitations to the use of CPC, where the *Porphyromonas gingivalis* only decreases with the use of CHX. Levels of periodontopathogens in CPC-treated biofilms were statistically similar to untreated biofilms. CPC only reduced 50% levels of *Porphyromonas gingivalis*.²¹ In addition, the concentration of CHX in CHX 0.2% was higher compared to the combination of CHX 0.12% and CPC 0.1%. These things allow the use of CHX 0.2% to be better than CHX 0.12% and CPC 0.1% against *Porphyromonas gingivalis*.

There are various other factors that affect the results of the differences in the inhibition zones formed. Factors that can affect the inhibition zone include the sensitivity of the organism, incubation temperature, incubation time, turbidity of the bacterial suspension, and thickness of the agar medium. The incubation temperature must be done at 37°C and for 24 hours, temperatures less than 37°C can cause a larger inhibition zone diameter. In bacterial suspension turbidity, the diameter of inhibition zone will be larger if the suspension is not more turbid than the turbidity of standard McFarland 0.5, and will be smaller if the suspension is more turbid. Less thickness of agar media can cause the diffusion process to be faster and vice versa.²²

CONCLUSION

Based on the results of the research, it can be concluded that CHX 0.2% mouthwash has greater inhibitory zone than CHX 0.12% and CPC 0.1% mouthwash against *Porphyromonas gingivalis*. There is a significant difference between the effectiveness of mouthwash containing CHX 0.12% and CPC 0.1% with CHX 0.2% mouthwash in inhibiting the growth of *Porphyromonas gingivalis*.

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REFERENCES

1. Lang NP, Bartold PM. Periodontal Health. J Periodontol. 2018; 89(1):9–16. DOI: 10.1002/JPER.16-0517.
2. Achmad H, Singgih MF, Huldani, Ramdhani AF, Ramadhany YF. Inhibitory Power Test of White Rice Bran Extract (*Oryza sativa L.*) with the Solution of Ethanol and Aquades on *Porphyromonas gingivalis* (In Vitro) Bacteria. Systematic Reviews in Pharmacy. 2020; 11(6):858–63. DOI: 10.31838/srp.2020.6.123.
3. Carranza FA, Newman MG, Takei HH, Klokkevold PR. Carranza's Clinical Periodontology. 13th ed. Philadelphia: Elsevier; 2019.
4. Mehrotra N, Singh S. Periodontitis. StatPearls Publishing. 2023.
5. Rafiei M, Kiani F, Sayehmiri F, Sayehmiri K, Sheikhi A, Azodi MZ. Study of Porphyromonas gingivalis in periodontal diseases: A systematic review and meta-analysis. Med J Islam Repub Iran. 2017; 31(62):2–5. DOI: 10.18869/mjiri.31.62.
6. Marsh PD, Lewis MA, Rogers H, Williams DW, Wilson M. Marsh and Martin's Oral Microbiology. 6th ed. Marsh and Martin's Oral Microbiology. UK: Elsevier; 2016.
7. Samaranyake L. Essential Microbiology for Dentistry. 5th ed. Essential Microbiology for Dentistry. Churchill Livingstone Elsevier; 2018.



8. Setiawatie EM, Valentina R, Meiliana RS. Effectiveness of Cetylpyridinium Chloride in Reducing the Growth of Bacteria that Cause Periodontal Disease. 2023; 11(2):115–20. DOI: 10.35790/eg.v11i.
9. Toar AI, Posangi J, Wowor V. Daya Hambat Obat Kumur Cetylpyridinium Chloride dan Obat Kumur Daun Sirih Terhadap Pertumbuhan *Streptococcus mutans*. Jurnal Biomedik. 2013; 5(1):163–8. DOI: 10.35790/jbm.5.1.2013.2639.
10. Takenaka S, Sotozono M, Ohkura N, and Noiri Y. Evidence on the Use of Mouthwash for the Control of Supragingival Biofilm and Its Potential Adverse Effects. Antibiotics. 2022; 11: 727.
11. Yarahmadi N, Hashemian F, Doust HH. Clinical Effects of Chlorhexidine 0.2% and Cetylpyridinium 0.05% Combination in Comparison with Chlorhexidine, Cetylpyridinium and Persica in Reducing Oral Bacteria in Healthy Individuals. J Pharm Care. 2020; 8(3):116–22. DOI: 10.18502/jpc.v8i3.4545.
12. Adkins KL. Effectiveness of cetylpyridinium chloride, chlorhexidine gluconate, chlorine dioxide, and essential oils against *F. nucleatum*, *P. gingivalis*, *S. mutans* and *S. sobrinus* - a biofilm approach. 2013; 5(1):15–23.
13. Hassan Najafi M, Taheri M, Reza Mokhtari M, Forouzanfar A, Farazi F, Mirzaee M, et al. Comparative study of 0.2% and 0.12% digluconate chlorhexidine mouth rinses on the level of dental staining and gingival indices. Dent Res J (Isfahan). 2013; 9(3):305–8.
14. Tartaglia GM, Tadakamadla SK, Connelly ST, Sforza C, Martín C. Adverse events associated with home use of mouthrinses: a systematic review. Ther Adv Drug Saf. 2019; 10(1):1–16. DOI: 10.1177/2042098619854881.
15. Dutt P, Kr Rathore P, Khurana D. Chlorhexidine - An antiseptic in periodontics. IOSR Journal of Dental and Medical Sciences. 2014; 13(9):85–8. DOI: 10.4103/jpbs.JPBS_162_20.
16. Sari D Novita, Cholil, Sukmana B Indra. Perbandingan Efektifitas Obat Kumur Bebas Alkohol Yang Mengandung Cetylpyridinium Chloride Dengan Chlorhexidine Terhadap Penurunan Plak. Dentino. 2014; 2(2):179–83.
17. Tille PM. Bailey & Scott's Diagnostic Microbiology. 13th ed. St. Louis, Missouri : Elsevier; 2014.
18. Sinaredi BR, Pradopo S, Wibowo TB. Daya antibakteri obat kumur chlorhexidine, povidone iodine, fluoride suplementasi zinc terhadap *Streptococcus mutans* dan *Porphyromonas gingivalis*. Dent J. 2014; 47(4):211–4. DOI: 10.20473/j.djmk.v47.i4.p211-214.
19. Setiawatie EM, Valentina R, and Meiliana RS. Effectiveness of Cetylpyridinium Chloride in Reducing the Growth of Bacteria that Cause Periodontal Disease. e-GiGi. 2023; 11 (2): 115-120 DOI: <https://doi.org/10.35790/eg.v11i2.44510>.
20. Mao X, Auer DL, Buchalla W, Hiller KA, Maisch T, Hellwig E, et al. Cetylpyridinium Chloride: Mechanism of Action, Antimicrobial Efficacy in Biofilms, and Potential Risks of Resistance. Antimicrob Agents Chemother. 2020; 64(8):2–4. DOI: 10.1128/AAC.00576-20.
21. De Miranda SLF, Damaceno JT, Faveri M, Figueiredo LC, Soares GMS, Bueno-Silva B. In Vitro Antimicrobial Effect of Cetylpyridinium Chloride on Complex Multispecies Subgingival Biofilm. Braz Dent J. 2020; 31(2):103–8. DOI: 10.1590/0103-6440202002630.
22. Zeniusa P, Ricky Ramadhian M, Hamidi Nasution S, Karima N. Uji Daya Hambat Ekstrak Etanol Teh Hijau Terhadap *Escherichia coli* Secara In Vitro. Majority. 2019; 8(2):136–43.



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