

Search

Home / Archives / Vol. 6 No. 12 (2022): Tropical Journal of Natural Product Research (TJNPR)

Published: 2022-12-30

Articles



A Review of the Phytochemical, Usability Component, and Molecular Mechanisms of *Moringa oleifera*
<http://www.doi.org/10.26538/tjnpr/v6i12.1>

Filza Y. Ade, Unang Supratman, Nesti F. Sianipar, **Julia W. Gunadi**, Putri Teesa Radhiyanti, Ronny Lesmana
1906-1913

PDF DOI



Development and Evaluation of a Topical Herbal Gel for the Treatment of *Tinea pedis*
<http://www.doi.org/10.26538/tjnpr/v6i12.2>

Erza Genatrika, Elza Sundhani, Mayang I. Oktaviana
1914-1918

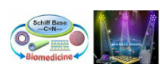
PDF DOI



Morphological Responses of *Cedrus atlantica*, *Pinus halepensis*, and *Tetraclinis articulata* in Different Pedoclimatic Conditions
<http://www.doi.org/10.26538/tjnpr/v6i12.3>

Safaa Serbouti, Younes Abbas, Meriem Soussi, Issam Alaoui, Wafae Squalli, Hamid Achiban
1919-1924

PDF DOI



Synthesis, Antibacterial, and Antioxidant Activities of Some Schiff Bases
<http://www.doi.org/10.26538/tjnpr/v6i12.4>

Hala Sabry Al-Atbi, Hassan A. Jaraf, Asmaa B. Sabti
1925-1929

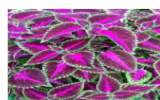
PDF DOI



Curcumin and Fluconazole to Resolve Fluconazole-resistant *Candida albicans* Infection in HIV Patient
<http://www.doi.org/10.26538/tjnpr/v6i12.5>

Novida Ariani, Mulyohadi Ali, Noorhamdani Noorhamdani, Aulanni'am Aulanni'am, Budi Santoso, Sumarno Sumarno, Bambang Rahardjo, Siti Chandra
1930-1935

PDF DOI



Platanus scutellarioides Leaf Extract Protective Effects Against Isoniazid and Rifampicin-Induced Hepatotoxicity in Wistar Rats
<http://www.doi.org/10.26538/tjnpr/v6i12.6>

Lusi P. Dwita, Ni Putu E. Hikmawanti, Juhairah Husniah, Muhammad Hazraj, Siti N. Bela, Eva Aryanti
1936-1940

PDF DOI



Identified Compounds from Ethyl Acetate Phase of Temu Mangga (*Curcuma mangga* Val.) Using LC-MS/MS and Their Potential as Anticancer Against MCF-7 Cells
<http://www.doi.org/10.26538/tjnpr/v6i12.7>



More Graphical Abstracts

Indexing & Abstracting



Tropical Journal of Natural Product Research

Q4 Analytical Chemistry
best quartile

SJR 2021
0.13
powered by scimagojr.com

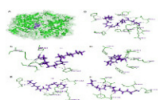
0.4 CiteScore²⁰²¹



<http://www.doi.org/10.26538/tjnpr/v6i12.7>

Alicia N.A. Ganur, Kurniawanti Kurniawanti, Purwantiningsih Sugita, Laksmi Ambarsari, Gustini Syahbirin, Auliya Ilmiawati, Dyah U.C. Rahayu
1941-1946

[PDF](#) [DOI](#)



Molecular Docking Discovered Potential of Cyclooxygenase – 2 Inhibitor Activity of Oily Compounds of Walnuts

<http://www.doi.org/10.26538/tjnpr/v6i12.8>

Sundari Sundari, Abdu Mas'ud, Dewi R. T. Sari
1947-1952

[PDF](#) [DOI](#)



The Protective Effect of Sa-khan (Piper, Piperaceae) Against High Glucose-Induced Cytotoxicity in Human Proximal Tubular Epithelial Cells
<http://www.doi.org/10.26538/tjnpr/v6i12.9>

Daoyot Daorueang
1953-1956

[PDF](#) [DOI](#)



Effects of Climate Change and Anthropization on Dynamics of Vegetation Cover in the Taounate Region of Morocco

<http://www.doi.org/10.26538/tjnpr/v6i12.10>

Abdelouahid Laftouhi, Noureddine Eloutassi, Elhachmia Ech-chihbi, Zakia Rais, Abdelfattah Abdellaoui, Abdeslam Taleb, Mustapha Beniken, Mustapha Taleb

[PDF](#) [DOI](#)



Leishmaniasis Risk Factors in Central Morocco: Urbanization and Socio-Economic Factors

<http://www.doi.org/10.26538/tjnpr/v6i12.11>

Hajar El Omari, Abdelkader Chahlaoui, Fatima Z. Talbi, Khadija Lahouiti, Abdelkarim Taam, Fatiha El Khayyat, Abdellatif Alami, Najoua Darkaoui, Abdelilah Merabti, Rachid Amaich, Abdelhakim El Ouali Ialami
1964-1968

[PDF](#) [DOI](#)



Statistical Indicators for Evaluating the Effect of Heavy Metals on Samaraa Drug Industry Water Exposed to the Sun and Freezing

<http://www.doi.org/10.26538/tjnpr/v6i12.12>

Layla R. Khaleel, Suadat M.M. Al-Hermizy, Mohammed N. Abbas
1969-1974

[PDF](#) [DOI](#)



Chemical Profiles and Bioactivities of Ethanol and Water Extracts of Sclerotium and Fruiting Body of Lignosus rhinocerotis (Cooke) from Banggai Islands, Indonesia

<http://www.doi.org/10.26538/tjnpr/v6i12.13>

Kris H. Timotius, Adit W. Santoso, Adelina Simamora
1975-1980

[PDF](#) [DOI](#)



Phytochemical, Antioxidant Analysis and In Vitro Xanthine Oxidase Inhibitory Activity of Kaempferia parviflora and Kaempferia galanga

<http://www.doi.org/10.26538/tjnpr/v6i12.14>

Tri B. Julianti, Mohd F.A. Bakar, Erindyah R. Wikantyasning
1981-1985

[PDF](#) [DOI](#)



Unique Marine Insects (Hemiptera) From North Sulawesi and Self-Defense Mechanism Against Sun Exposure

<http://www.doi.org/10.26538/tjnpr/v6i12.15>

Veibe Warouw, Joice R.T.S.L Rimper, Silvana D. Harikedua
1986-1989

[PDF](#) [DOI](#)



Phytochemical Analysis of Quercetin, Catechol and Tannic Acid in Ethanol Extract of Barleria Prinniflora Leaf by RP-HPLC Technique

18th percentile
Powered by **Scopus**

Keywords

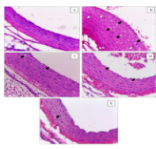




Extract of *Annona muricata* Linn Leaf by HPLC Techniques
<http://www.doi.org/10.26538/tjnpr/v6i12.16>

Soumyajit Das, Bannimath Gurupadayya, Hemanth P. R Vikram, Ittagi Shanmukha, Aparajita Neogi, Bannimath Namitha
 1990-1999

PDF DOI



Black Cumin Inhibits Pro-Atherogenic Changes and Reduces Aortic Intima Media Thickness in Rats with Sub-Chronical Cigarette Smoke Exposure
<http://www.doi.org/10.26538/tjnpr/v6i12.17>

Fita Triastuti, Meity Ardiana, Andrianto Andrianto, Hanestya O. Hermawan, Dara N. Ghassani
 2000-2006

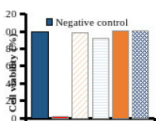
PDF DOI



In vitro Antioxidant and Immunological-Associated Activities of Ethanol Extracts of *Azima sarmentosa* (Blume) Benth. & Hook. F
<http://www.doi.org/10.26538/tjnpr/v6i12.18>

Neeranuch Sankla, Panida Loutchanwoot, Surasak Khankhum, Saranyu Khammuang, Rakrudee Sarnthima, Nuchsupha Sunthamala
 2007-2013

PDF DOI



Development of Topical Formulations with Antibacterial Efficacy from Peel Extract of *Mangifera indica*: Emulsions and Micro-emulsions
<http://www.doi.org/10.26538/tjnpr/v6i12.19>

Pornsak Sriamornsak, Arzu Polat, Bighan Torabi, Wantanwa Krongrawa, Anne Krüger-Genge, Joachim Storsberg, Tassilo Seidler, Mont Kumpugdee-Vollrath
 2014-2019

PDF DOI



Repellent Activity of Lotion of Essential Oils from *Piper betle* Linn and *Cymbopogon nardus* [L.] Rendle
<http://www.doi.org/10.26538/tjnpr/v6i12.20>

Rasidah Rasidah, Icha S. Maurizka, Vonna Auliashah, Munira Munira, Noni Zakiah, Rini Handayani
 2020-2024

PDF DOI



Identification of Active Antioxidant Compounds with Neuro-protective Effects in Kora-kora Coastal Macro-algae
<http://www.doi.org/10.26538/tjnpr/v6i12.21>

Finy Warouw, Christi Mambo, Junita M. Pertiwi, Veibe Warouw, Robert A. Bara
 2025-2028

PDF DOI



Effect of *Anchomanes difformis* Ethanol Root Extract on Electrolyte Level of Testosterone-Induced Benign Prostatic Hyperplasia in Wistar Rats
<http://www.doi.org/10.26538/tjnpr/v6i12.22>

Ukpanukpong R. Undigweundeye, Igbineweka P. Osagie, Eban L. kechi, Omang W. Achu, Patrick E. Bernard, Eteng U Mbeh
 2029-2034

PDF DOI



Elucidation of the Phytochemicals, Safety Profile, and Preclinical Anti-Inflammatory Activity of Ethanol Extract of *Combretum paniculatum* Leaves
<http://www.doi.org/10.26538/tjnpr/v6i12.23>

Ifeoma F. Chukwuma, Kelechi T. Aniagboso, Blessing E. Atrogo, Ukinebo E. Ese, Obasi Ezeali, Anthony U. Eze, Sandra N. Onuorah, Stanley T. Onyishi, Benita M. Titus, Emmanuel I. Ugwu
 2035-2040

PDF DOI



ACL-4, an Endophytic Fungus Isolated from *Ageratum conyzoides* Leaves Possesses the Unique Potential of Generating Low Molecular Weight Bioactive Lead Compounds
<http://www.doi.org/10.26538/tjnpr/v6i12.24>

Chinwuba O. Obidiegwu, Chika C. Abba, Nkeoma N. Okoye, Chukwubikem C. Okolo, Charles U. Nwachukwu, Nonye T. Ujam, Festus B. C. Okoye

2041-2046

[PDF](#) [DOI](#)

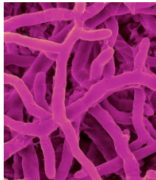


Antihypertensive and Cardioprotective Effects of *Salvia officinalis* (Sage) Leaf in N^GNitro-L-Arginine Methyl Ester (L-NAME) Induced-Hypertensive Wistar Rats

<http://www.doi.org/10.26538/tjnpr/v6i12.25>

Azuka T.H. Mokogwu, Collins O. Adjekuko, Uzoh H. Oshilonyah, Joan O. Ikpefan, Godwin O. Awwioro
2047-2050

[PDF](#) [DOI](#)



Antifungal Activities of Black Seed, Castor and Lemon Oils Against Pathogenic Plant Fungi

<http://www.doi.org/10.26538/tjnpr/v6i12.26>

Idowu A. Obisesan, Emmanuel O. Tubi, Oluwatomi O. Eyinade, Oluwakemi M. Adeniran
2051-2056

[PDF](#) [DOI](#)

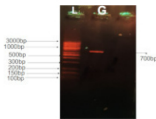


Comparative Studies on Anti-Diabetic Properties of Commonly Consumed *Musa* Cultivars (*M. paradisiaca*, *M. sapientum*, and AAB Group Hybrid) Flour

<http://www.doi.org/10.26538/tjnpr/v6i12.27>

Ganiyu Oboh, Agunde S. Oluwakemi, Ademosun O. Ayokunle, Idowu S. Oyeleye
2057-2062

[PDF](#) [DOI](#)



Kinetics and Thermodynamic Properties of Pectinase Obtained from *Trichoderma longibrachiatum* MT321074

<http://www.doi.org/10.26538/tjnpr/v6i12.28>

Chukwunonso A. Nsude, Tobechukwu C. Ezike, Arinze L. Ezugwu, Ozoemena E. Eje, Ikechukwu N. E. Onwurah, Ferdinand C. Chilaka
2063-2072

[PDF](#) [DOI](#)

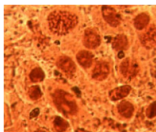


Assessment of Pharmacodynamic Interactions and Toxicological Effects of *Vernonia amygdalina* –Metformin Co-Administration on Streptozotocin-Induced Diabetic Wistar Rats

<http://www.doi.org/10.26538/tjnpr/v6i12.29>

Michael A. Olamoyegun, Nusrat O. Omisore, Adekemi O. Yusuf, Tola E. Odewale
2073-2080

[PDF](#) [DOI](#)



Cytotoxicity Evaluation and the Free Radical Scavenging Activity of the Root Extracts of *Calotropis procera*

<http://www.doi.org/10.26538/tjnpr/v6i12.30>

Godwin O. Ihegboro, Chimaobi J. Ononamadu, Tajudeen A. Owolarafe, Rachael Kolawole, Mujiburrahman Fadilu, Abubakar M. Aminu, Emmanuel Afor, Sandra N. Ihegboro, Muhamud Adebayo, Yunusa Abdullahi
2081-2086

[PDF](#) [DOI](#)

ABOUT TJNPR

[About the Journal](#)
[Aims & Scope](#)
[Editor Profile](#)
[Editorial Board](#)

RESOURCES

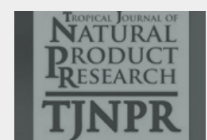
[Editorial Policy & Malpractice Statement](#)
[Guide for Authors](#)
[Guidelines for Reviewers](#)
[Open Access Policy](#)

MY ACCOUNT

[Authors](#)
[Editors](#)
[Reviewers](#)
[Subscribers](#)

ISSUES

[Current](#)
[Archives](#)



**A Review of the Phytochemical, Usability Component, and Molecular Mechanisms of *Moringa oleifera***Filza Y. Ade¹, Unang Supratman³, Nesti F. Sianipar^{4,5}, Julia W. Gunadi⁶, Putri Teesa Radhiyanti^{2*}, Ronny Lesmana^{1,2}¹ Biological Activity Division, Central Laboratory, Universitas Padjadjaran, Bandung, 45363, Indonesia² Physiology Division, Department of Biomedical Sciences, Faculty of Medicine, Universitas Padjadjaran, Bandung, 45363, Indonesia³ Departement of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Padjadjaran, Bandung, 45363, Indonesia⁴ Food Technology Department, Faculty of Engineering, Bina Nusantara University, Jakarta, 11480, Indonesia⁵ Research Interest Group Food Biotechnology, Bina Nusantara University, Jakarta, 11480, Indonesia⁶ Department of Physiology, Faculty of Medicine, Maranatha Christian University, Surya Sumantri 65, Jawa Barat 40164, Indonesia

ARTICLE INFO

ABSTRACT

Article history:

Received 06 June 2022

Revised 16 November 2022

Accepted 02 December 2022

Published online ****

Copyright: © 2022 Ade *et al.* This is an open-access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Moringa oleifera (*M. oleifera*) or kelor is a well-known herbal plant. *M. oleifera* is believed to be a magical tree because it provides tremendous benefits and is a rich source of nutrients for all living things. The potential value of *M. oleifera* in preventing or treating various chronic diseases is enhanced. There are still scattered details about phytochemical and usability components and their molecular mechanisms. The purpose of this article is to collect the various scattered data to make it easier for readers to get the information. Thus, in this study, we discussed information comprehensively for the phytochemical components of *M. oleifera* and the potential benefits of biological activities. This review will give information to open the possible utilization of *M. oleifera*, especially for the patient's medication and/or supplementation.

Keywords: Biological activities, Molecular mechanisms, *Moringa oleifera*, Phytochemical Components, Plant.

Introduction

Moringa oleifera, also called Kelor, is one of the most well-known herbal plants. *Moringa oleifera* is one of the most commonly used medicinal herbs in the *Moringa* genus. The *Moringa* plant is native to India's southern Himalayan foothills and has been cultivated in almost every tropical area under different names depending on the region. *Moringa* is known as the Drumstick Tree in English because of its drumstick-shaped pods, but it is also known as the Miracle Tree or Magic Tree¹ due to its various health benefits. It has a wide variety of medicinal properties, and a high nutritional value,^{2,3} a source of nutrients that can prevent malnutrition,⁴ such as protein, vitamins, minerals, and amino acids,^{2,5} and it cures various diseases.⁶

Moringa plants are relatively easy to grow and disseminate, both sexually and asexually, and do not need a lot of nutrients or water, making it simple to manage production on a large or household scale. It can also help to improve environmental soil conditions. *Moringa* is a plant that can withstand a wide range of environmental conditions, including diverse climates, lousy soil conditions, and average dryness.⁷ This plant grows well in areas with adequate lighting or sunlight exposure and thrives at an altitude of 600m - 2000m a.s.l, reaching a height of 152.40 cm in 5-6 months. The *Moringa* plant is a tree with a height of up to 10-12 m,⁸ woody roots, and thin skins that can thrive in highland and lowland environments, with a generative (seed) or vegetative (stem cuttings) distribution. In sandy or loamy soils, the temperature for growth varies from 25 to 35°C (77-95 °F), and it is very tolerant of clay soils.⁷

Plant growth is affected by variations in environmental factors such as geographic location, altitude, and temperature/climate where it grows. Furthermore, genetic variation in a plant affects the nutritional content and phytochemical composition of each part of the plant, Cultivar conservation and growth of breeding programs to grow superior cultivars,⁹ genetic explorations, as well as genetic improvement initiatives,¹⁰ and the future selection and breeding programs aimed at tree improvement¹¹, can all benefit from genetic diversity studies. *Moringa* has a wide range of characteristics and morphological variability, which can be used to conserve and identify *M. oleifera* germplasm.⁷

Moringa oleifera is rich in various nutrients such as carotenoids, tannins, flavonoids, anthraquinones, steroids, alkaloids, terpenoids, anthocyanins, glycosides, and saponins¹² which are abundant in different parts of the *M. oleifera* plant. Amino acids, flavonoids, phenolic acids, vitamins, glucosinolates, carotenoids, tannins, alkaloids, polyphenols, isothiocyanates, and saponins are available in the plant's leaves.^{4,7} The leaves of *M. oleifera* are rich in vitamins, minerals, and various health-promoting secondary metabolites. The high concentration of bioactive compounds in *M. oleifera* leaves accounts for their pharmacological properties.⁷ The *Moringa* plant's phytochemicals help prevent diabetes, inflammation, cancer, arthritis, cardiovascular disease, and age-related functional disorders, among other health problems. *M. oleifera* has been shown to have anti-hypertensive, anti-diabetic,² anti-inflammatory, anti-cancer,⁴ and antioxidant potential.^{4,5} *M. oleifera's* anticancer, antioxidant, antimicrobial, and anti-diabetic effects are all due to its bioactive compounds.¹² Extracts of the leaves of *Moringa* (*Moringa oleifera*) have different phytochemical content, which showed a significant benefit in herbal medicine, one of them is in the anti-obesity activity.¹³ The pathogenesis of obesity involves the accumulation of lipids, the formation of oxidative stress, and inflammatory reactions. As one of the anti-obesity agents, *Moringa's* leaf extract has antioxidant, anti-inflammatory, and hypolipidemic effects. *Moringa* leaf extract contributes to modifying lipid metabolism in the mevalonate pathway.¹³ The mevalonic pathway is one of the pathways that involve

*Corresponding author. E mail: putri@unpad.ac.id
Tel: +62 22 7795594

Citation: Ade FY, Supratman U, Sianipar NF, Gunadi JW, Radhiyanti PT, Lesmana R. A Review of the Phytochemical, Usability Component, and Molecular Mechanisms of *Moringa oleifera*. Trop J Nat Prod Res. 2022; 6(12):

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

adipogenesis.¹⁴ Adipogenesis inhibition is crucial for developing anti-obesity drugs, and it is a significant field of research. Astragaloside increases lipolysis and suppresses adipogenesis in 3T3-L1 adipocytes, making it a promising candidate for obesity treatment.¹⁵

Methods

The review was based on studies discovered using the keywords *Moringa oleifera*, phytochemical component, the usefulness of *Moringa oleifera*, genetic diversity, *Moringa* molecular mechanism, and other similar terms in electronic databases such as Google Scholar, Elsevier, PubMed, and Mendeley. Unrelated studies were excluded. We gathered all of the relevant research papers. The literature method diagram is shown in Figure 1.

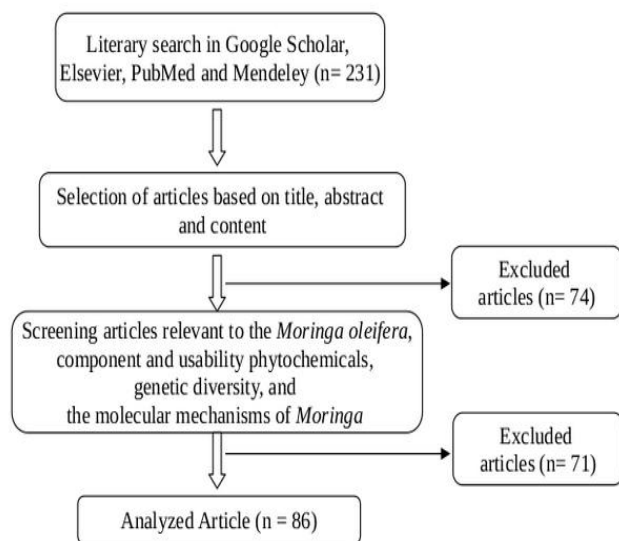


Figure 1: Method diagram of literature review

Results and Discussion

Moringa oleifera Phytochemical Material

Research on the phytochemical content of each part of the *Moringa* plant has also been carried out and is presented in Table 1. The leaves contain amino acids (arginine, histidine, methionine, cysteine, tryptophan, and lysine),^{5,25} vitamins (A, B, B1, B2, B3, C),^{5,25,26} minerals (Mg, Ca, K, phosphorus, sodium, sulfur, zinc, copper, manganese, iron, selenium),^{5,21,25} alkaloids,¹⁸ phenolic acids,^{5,16,18,21,25,27,28,29} flavonoids,^{5,16,18,24,27,28,29} terpenoids,¹⁷ polyphenols,^{19,24,25} ascorbic acid,²¹ protein, folic acid, β -sitosterol, iron, α -tocopherol, oestrogenic substances, riboflavin, β -carotene, nicotinic acid, pyridoxine,^{25,26,38} thiocarbamate glycosides, nitrile, and mustard oil glycosides.^{20,22,23} Those components have antioxidant activity,^{5,16,18,19,24,28,29,36} antiinflammatories,^{5,16} antibacterial,^{5,16} antimicrobials,^{30,36} anticancer,⁵ antidiabetic,^{5,24} and antihyperglycemic effects.¹⁷ The leaves virtually are ideal for dietary supplements²⁶ and responsible in lowering the blood pressure.^{20,22,23} *Moringa* leaves contain mainly phenolic compounds, which are natural plant-based micronutrients. Flavonoids and tannins are among the several forms of polyphenols. Kaempferol and quercetin derivatives are primarily composed of flavonoids.¹³ The presence of β -sitosterol, 4-hydroxymellin, β -citostenone, vanillin, octacosanoic acid,²⁰ phenolic, alkaloids,³¹ which have potential antimicrobial activity,^{30,36} antioxidant,^{31,36} are found in the bark of moringa. Meanwhile, in the *Moringa* seeds, there are ash, fat, seven bioactive compounds, carbohydrates, β -sitosterol-3-O- β -D-glucopyranoside, 4 (α -L-rhamnosyloxy)-benzyl isothiocyanate, niazirin, 3-O-(6'-O-oleoyl- β -D-glucopyranosyl)- β -sitosterol, glycerol-1-(9-octadecanoic), β -sitosterol, vitamin,³³ niazimicin,² flavonoids,³² ascorbic acid, protein,^{33,38} and has antimicrobial activity,³⁰ antitumors,^{2,}

³³ antioxidants,³² and anti-obesity.³⁴ In *Moringa* flowers were found the presence of amino acids, kaempferol, sucrose, wax, D-glucose, alkaloids, quercetin,³⁵ potassium, calcium, flavonoid pigments (alkaloids, kaempferitrin, rhamnetin, isoquercitrin, kaempferol), vitamin, and ascorbic acid.^{20,31} The flower's components were used as antioxidant and diuretic agent.³¹ Diuretic activities eventually will stabilize the blood pressure.³¹ In the *Moringa* root, alkaloids³⁶ and protein³⁸ are found, which have the potential for antimicrobial activity,^{30,36} diuretic action, antioxidant,³⁶ and anti-inflammatory.³⁷ Isothiocyanate glycosides, thiocarbamate,³⁹ ascorbic acid, phosphorus, oestrogenic substances, iron, riboflavin, β -sitosterol, pyridoxine, protein, calcium, vitamins, nicotinic acid, β -carotene, α -tocopherol, amino acids (methionine, cystine, tryptophan, and lysine), copper, and folic acid;³² are found in *Moringa* pods and have potential as an antioxidant^{2,40,41} and an antihypertensive.³⁹ The chemical structure figures of the *Moringa*'s seed, pod, root, bark, leaf, and flower compounds are presented in figure 2.

Uses of *Moringa oleifera*

Every part of the *Moringa* plant (roots, stems, leaves, flowers, and fruit) has been widely researched and used in medicine, food ingredients, dyes, animal feed, water purification, and renewable energy source for ethanol raw materials. The *Moringa* plant contains many beneficial chemicals.³⁶ *Moringa* leaves, roots, and seeds contain essential medicinal and antioxidant components such as alkaloids, phenolic acids, flavonoids, and tannins.⁴³⁻⁴⁹ In addition, research on the nutrients contained in *Moringa* plants has also been carried out, including Bharali et al. (2003) stating that *Moringa* plants contain pro-vitamins A and C, especially β -carotene, which will be converted into vitamins in the body;⁵⁰ Saini et al. (2016) found vitamins A, C, calcium, protein in *Moringa* leaves. The presence of gluconate and isothiocyanate compounds found in *Moringa* plants can be an anticancer agent and inhibit bacterial and fungal activity;² *Moringa* oil contains oleic acid, which can be a raw material for biodiesel.⁵¹ Research on the medicinal properties of *Moringa* has also been carried out, including the anti-tumor activity,^{33,52} antimicrobial,³⁶ anticancer,^{2,53} allergies,⁴² antioxidants,^{24,28,31,36,40,49,53,54,55} anti-inflammatories,^{37,53} anti-obesity,³⁴ antihyperglycemics (antidiabetic).^{24,34} Therefore, *Moringa* is commonly used as a phytotherapeutic agent in Africa and India.⁵⁶ Interest in research on *Moringa* and its medicinal properties and health, as well as on the utilization of the environment such as water purification, conservation, breeding, and increasing the variability of the species, are genetical.^{7,57,58,59,60,61}

Molecular Mechanism of *Moringa*

The plant *Moringa oleifera*, which has benefits and nutritional and medicinal value, has been widely reported in various studies regarding the potential activity and molecular mechanisms of the extract of every part of the *Moringa* plant. *M. oleifera* extracts have been shown to have a variety of nutraceutical and pharmacological properties, including anti-inflammatory properties,⁴³ anti-oxidant,^{52,62} anti-obesity,^{62,63} anti-cancer,⁴³ anti-tumor, hepatoprotective,⁵² neuroprotective, hypoglycemic, anti-diabetic, and blood fat reduction.^{64,65,66} Some studies in the treatment of diabetes used *M. oleifera* leaves, which have many dissolved compounds involved in glucose homeostasis, including isothiocyanates which can reduce insulin resistance and liver gluconeogenesis.⁶⁷ *Moringa* leaves contain many polyphenolic compounds, such as flavonoids and phenolic acid, which may also play a role in glucose homeostasis, which exerts anti-diabetic effects.⁴³ The glucose homeostatic effect may also be due to anti-oxidants, receptor agonists or antagonistic activity, and enzyme inhibition.⁶⁴⁻⁶⁶ Although several scientific studies on *Moringa* leaves' hypoglycemic effect have been conducted, more information about age, gender, nutritional status, ethnicity, and dietary habits in humans is required before using the leaves as a herbal medicine for diabetes treatment. Compared to *Moringa stenopetala* leaf, the *Moringa oleifera* leaf has hypolipidemic activity, suggesting that its extract could be used in obesity management.⁶⁸ Latest study revealed that *Moringa oleifera* leaf extract could hinder some enzymes, such as pancreatic lipase and pancreatic cholesterol esterase. This study also showed that *Moringa oleifera* leaf extract prevents cholesterol

micellization formation. *Moringa* leaf contains chlorogenic acid, which inhibits lipogenesis and cholesterol production.⁶⁹

Table 1: Phytochemicals and usability components of *Moringa oleifera*

Plant Part	Phytochemical components	Medicinal uses / potential activity
Leaf	amino acid ^{5,25} , vitamin ^{5,25,26} , mineral ^{5,21,25} , phenolic ^{5,16,18,21,25, 27, 28, 29} , flavonoid ^{5,16,18,24,27,28,29} , Terpenoid ¹⁷ , Polyphenols ^{19,24,25} , alkaloid ¹⁸ , ascorbic acid ²¹ , folic acid, β -sitosterol, iron, α -tocopherol, oestrogenic substances, riboflavin, β -carotene, nicotinic acid, pyridoxine, protein ^{25,26,38} , thiocarbamate glycosides, Nitrile, and mustard oil glycosides ^{20,22,23}	Antioxidants ^{5,16,18,19,24,28,29,36} , antimicrobial ^{30,36} , antidiabetic ²⁴ , anti-inflammatory ^{5,16} , antibacterial ^{5, 16} , antihyperglycemic ¹⁷ , virtually ideal dietary supplement ²⁶ , responsible for the reduction of blood pressure ^{20,22,23}
Bark	Alkaloid (moringine, moringinine) ³⁶ , β -sitosterol, 4-hydroxymellin, β -citostenone, vanillin, octacosanoic acid ²⁰ , phenolic ³¹	Antimicrobial ^{30,36} , antioxidant ^{31,36}
Pods	isothiocyanate glycosides, thiocarbamate ³ , ascorbic acid, phosphorus, estrogenic substances, iron, riboflavin, β -sitosterol, pyridoxine, protein, calcium, vitamins, nicotinic acid, β -carotene, α -tocopherol, amino acids (methionine, cystine, tryptophan and lysine), copper, folic acid ³²	Antioxidant ^{2,40,41} , Antihypertensive ³⁹
Seed	Flavonoid ³² , β -sitosterol-3-O- β -D-glucopyranoside, 4 (α -L-rhamnosyloxy)-benzyl isothiocyanate, niazirin, 3-O-(6'-O-oleoyl- β -D-glucopyranosyl)- β -sitosterol, glycerol-1-(9-octadecanoic), β -sitosterol, ³³ niazimicin ^{2,33} , vitamin, ascorbic acid, protein ³⁸	Antimicrobial ³⁰ , antitumor ^{2,33} , antioxidant ³² , stabilizing effect on blood pressure, anti-obesity ³⁴
Flower	nine amino acids, kaempferol, sucrose, wax, D-glucose, traces of alkaloids, quercetin ³⁵ ; potassium and calcium, flavonoid pigments (alkaloids, kaempferitrin, rhamnetin, isoquercitrin, kaempferol), vitamin, ascorbic acid ²⁰ .	Diuretic activities, stabilizing effect on blood pressure, antioxidant ³¹
Root	Alkaloids ³⁶ , protein ³⁸	Antimicrobial ^{30,36} , diuretic activities, antioxidant ³⁶ anti-inflammatory ³⁷

Moringa oleifera leaf extract (MLE) has been shown to inhibit lipogenesis in 3T3-L1 adipocytes via the 5' adenosine monophosphate-activated protein kinase (AMPK) pathway. AMPK is a major glucose and lipid metabolism regulator at the cellular level. AMPK is a major glucose and lipid metabolism regulator at the cellular level.¹³ The lipolysis of 3T3-L1 adipocytes was improved by quercetin. Chlorogenic acid's reduction of insulin in MLE indicated that insulin sensitivity was improving positively.⁶⁹ Insulin sensitivity was improved by kaempferol. In MLE, quercetin and kaempferol had the most significant potential for binding to α -glucosidase.⁷⁰ In 3T3-L1 cells, kaempferol potency, was also found to have anti-adipogenic properties in a previous study.⁷¹ MLE treatment improved lipid profiles in 3T3-L1 adipocytes.⁷⁰ In high-lipid rats, astragaloside, the 3-O-glucoside form of kaempferol, showed anti-hyperlipidemic activity.¹³ In 3T3-L1 adipocytes, these anti-adipogenic effects could reduce leptin gene expression and secretion. The protein pro-lipogenesis sterol regulatory element binding proteins (SREBP) transcription factor had been an inhibitory target of MLE phytochemistry, astragaloside.¹⁵ SREBP was essential for activating the SREBP/mevalonate pathway linked to lipid metabolism. Yes-associated protein (YAP) and transcriptional coactivator with PDZ-binding motif (TAZ) activity, also called YAP/TAZ activity, was promoted by the SREBP-mevalonate axis.¹³ MLE has also been shown to block the HMG-CoA reductase enzyme.⁷² The presence of SREBP and HMG-CoA reductase in the mevalonate pathway has been a priority of MLE in reducing adipogenic activity.¹⁵ Statins have been shown in vitro to inhibit mevalonate pathways and YAP/TAZ activity.⁷³ Several studies have implicated the Hippo-YAP / TAZ

pathway in organ size regulation, tissue regeneration, organ formation, and self-renewal.^{73,74} Thus, the Hippo-YAP / TAZ line is one of the central components in network homeostasis.⁷⁴ Quercetin and kaempferol in MLE have been shown to inhibit PPAR γ activity in previous studies. YAP/TAZ signaling was also found to control PPAR γ .¹³ PPAR γ continues to express adipogenic genes when carrying out its transcription function.⁷⁵ Along with their extraordinary biological properties in growth, cancer, and tissue homeostasis, the transcription regulators Yes-associated Protein (YAP) and Transcriptional Co-activator with PDZ-binding motif (TAZ) are the main focus of attention.⁷⁶ The mevalonate pathway regulates YAP / TAZ action.⁶⁹ The YAP and TAZ read various mechanical cues, including shear stress, cell structure, and extracellular matrix stiffness, and convert them into cell-specific transcription programs. Mechanotransduction in YAP and TAZ is needed for behavior and stem cell regeneration. They provide new insights into how aberrant cell processes contribute to developing diseases such as inflammation, atherosclerosis, muscular dystrophy, fibrosis, cancer, and pulmonary hypertension.⁷⁷ The Hippo pathway, which controls homeostasis and plays a crucial role in carcinogenesis and regenerative processes, is one of the most critical cellular signaling pathways.⁷⁸ YAP/TAZ signaling has been shown to affect lipid metabolism and insulin sensitivity. In adipose-derived human mesenchymal stem cells treated with MLE, the expression of the insulin receptor substrate (IRS1) gene increased.⁷⁹ The TAZ protein influenced glucose absorption by GLUT4 by upregulating IRS1.⁸⁰ *M. oleifera* Lam. It ultimately uses thermogenic pathways to influence lipid metabolism. In a previous study,⁷⁹ uncoupling protein 1 (UCP1) was overexpressed

as a thermogenesis marker during adipose tissue differentiation. UCP1 control is inextricably linked to YAP/TAZ signaling.⁸¹

YAP and TAZ are Hippo signaling pathway downstream effectors that play a crucial role in cancer growth, including metastasis.⁸²

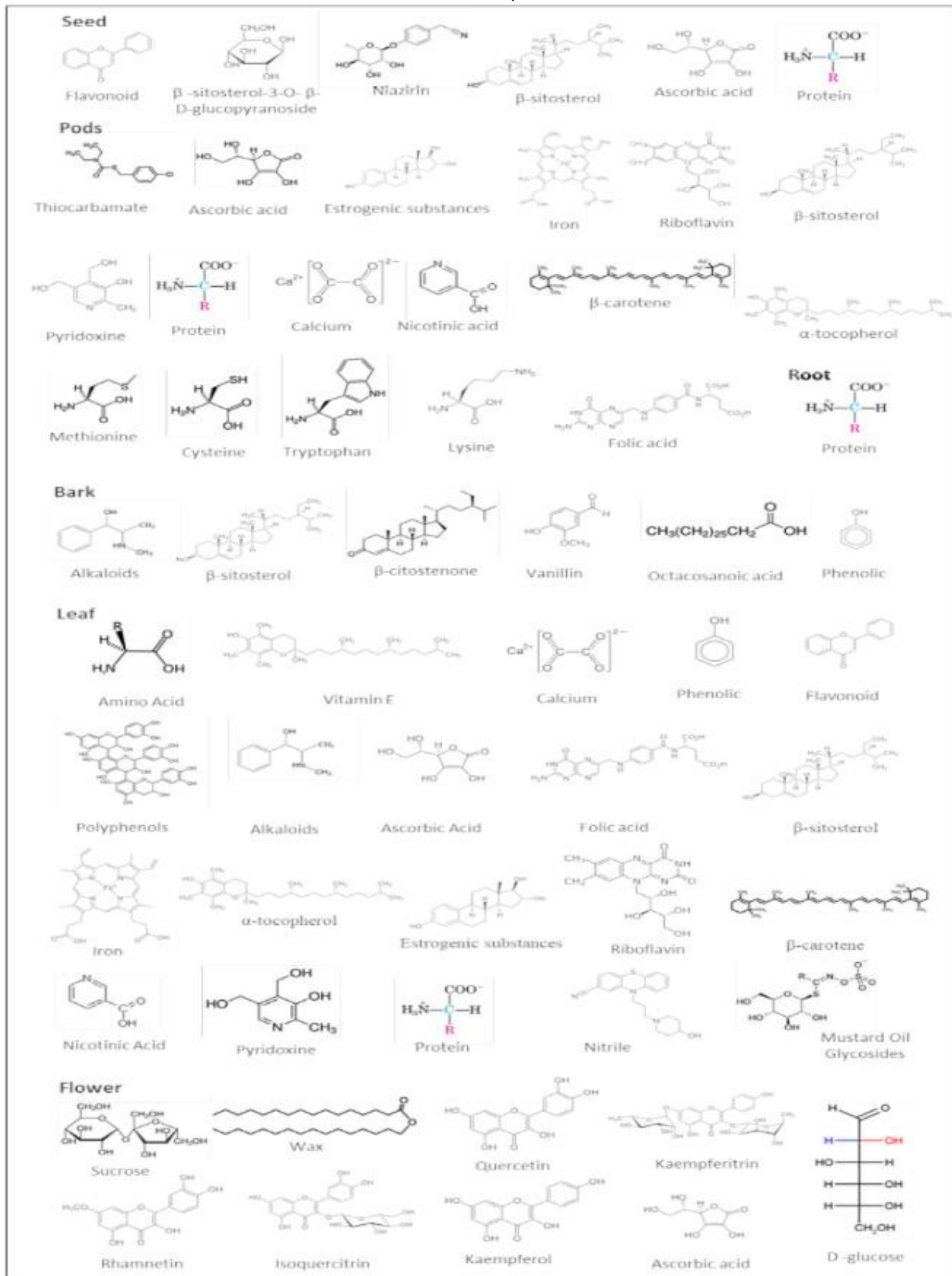


Figure 2: Chemical Structure of Moringa's Compound

Cell adhesion and mechanical signals had received from the network architecture and surrounding extracellular matrix (ECM) is also modulated by YAP/TAZ activation.⁷⁶ The Hippo signaling pathway regulates various physiological processes, and its dysfunction has increased the number of human diseases, including cancer.⁸³ YAP/TAZ controls cell metabolism in response to phenotypic changes, and recent research has shown that metabolic changes induced by YAP/TAZ lead to metastasis.⁸² *M. oleifera* leaves contain various beneficial natural substances such as flavonoids, ascorbic acid, phenolic acids, and carotenoids, making them an excellent antioxidant source.⁸⁴ In the variation of antioxidant activity, the primary and lateral roots of *M. oleifera* had higher antioxidant activity than the leaves.⁸⁵ Isothiocyanates, glucosinolates, and thiocarbamates were phytochemical constituents with significant antioxidant activity from the *M. oleifera* plant.⁸⁶ Antioxidant elements in *M. oleifera*, such as phenolics, flavonoids, ascorbic acid, and carotenoids, have been shown to increase the shelf life of lipid foods.⁵ Another research has shown that *M. oleifera* seed oil containing oleic acid has a significant anti-inflammatory effect by interfering with PKC pathways linked to the glucocorticoid response.⁸⁷ Flavonoids, phenols, tannins, saponins, vanillin, β -sitosterol, and terpenoid present in MLE were thought to be responsible for all these anti-inflammatory activities.⁸⁸ While quercetin and β -sitosterol have been linked to MLE's hypolipidemic activity, they also have anti-inflammatory properties. Due to their pleiotropic effects, both compounds restored normal hepatic morphology and physiology.⁸⁹ The molecular mechanism of MLE's effect on autophagy has been approached slowly in research. Autophagy is the degradation of a specific cell component or cytoplasmic substance within the cell.⁹⁰ In a study on MLE's antioxidant and anti-inflammatory effects through the autophagic route, isothiocyanates (ITCs) were highlighted. β -sitosterol has similar anti-inflammatory properties in vitro.^{91,92} *M. oleifera* is an effective antibacterial agent, as shown by a study that found a significant reduction in the growth of test bacteria.⁹³ The antibacterial and antifungal activities of the aglycone of deoxy-niazimicin obtained from the chloroform fraction of ethanolic extracts were found in the root bark of *M. oleifera*.⁴³ Aqueous and ethanol extracts of *M. oleifera* showed strikingly close results in lowering blood pressure in another study.⁸⁴ *M. oleifera* saponin has been reported as having antihypertensive properties.⁹⁴ The substance responsible for these effects, like thiocarbamate, and isothiocyanate glycosides, was successfully separated from the acetate stage of the ethanol extract of *M. oleifera* pods in a study that further supports the antihypertensive behavior of *M. oleifera* pods.⁸⁴ In many studies, *Moringa oleifera* leaves have been shown to have anticancer properties.⁹⁴ Isothiocyanate and thiocarbamate are two related bioactive compounds that have been isolated and have an inhibitory effect on the tumor promoter.⁴² Another research⁹⁵ found that *M. oleifera* leaf extract in an aqueous medium decreased the presence of cancer cells in the pancreas, tumor formation, and metastatic activity. *M. oleifera* exhibited cell death via the induction of apoptosis and necrosis mechanisms.⁹⁶ *Moringa oleifera* can also improve the killing of cervical cancer cells.⁹⁷ Another study⁹⁸ found that *Moringa* leaves suppressed AOM/DSS-induced colorectal carcinogenesis in a vivo model, suggesting that *Moringa*'s phenolics and complete dietary fiber may have chemopreventive potential. All the results presented in this review can ideally serve as a foundation for future research into the molecular mechanism of *M. oleifera*.

Conclusion

Moringa oleifera is a rich source of nutrition and has the potential value in preventing or treating several chronic diseases in the last few years. Many literature reviews had shown the utilization of various parts of *Moringa oleifera* in biology medicine and pharmacology. Although many studies in *Moringa oleifera*, unfortunately, details investigation and analysis into this plant are not well reported. There is an expectation to provide protocols and medicinal properties or supplementation for patients' treatment.

Conflict of Interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

Acknowledgments

This work was supported by funding from Hibah Post-Doctoral Universitas Padjadjaran (3570/UN6.3.1/LT/2020) to FYA.

References

- Bartlett DB, Willis LH, Slentz CA, Hoselton A, Kelly L, Huebner JL, Kraus VB, Moss J, Muehlbauer MJ, Spielmann G, Kraus WE, Lord JM, Huffman KM. Ten weeks of high-intensity interval walk training is associated with reduced disease activity and improved innate immune function in older adults with rheumatoid arthritis: A pilot study. *Arthritis Res Ther*. 2018; 20(1):1-15.
- Bartlett DB, Slentz CA, Willis LH, Hoselton A, Huebner JL, Kraus VB, Moss J, Muehlbauer MK, Spielmann G, Muoio DM, Koves TR, Wu H, Huffman KM, Lord JM, Kraus WE. Rejuvenation of Neutrophil Functions in Association With Reduced Diabetes Risk Following Ten Weeks of Low-Volume High Intensity Interval Walking in Older Adults With Prediabetes – A Pilot Study. *Front Immunol*. 2020; 11:1-14.
- Flack KD, Davy BM, Deberardinis M, Boutagy NE, McMillan RP, Hulver MW, Frisard MI, Anderson AS, Savla J, Davy KP. Resistance exercise training and in vitro skeletal muscle oxidative capacity in older adults. *Physiol Rep*. 2016; 4(13):1-8.
- Nyberg M, Blackwell JR, Damsgaard R, Jones AM, Hellsten Y, Mortensen SP. Lifelong physical activity prevents an age-related reduction in arterial and skeletal muscle nitric oxide bioavailability in humans. *J Physiol*. 2012; 590(21):5361-5370.
- Orlando P, Silvestri S, Galeazzi R, Antonicelli R, Marcheggiani F, Cirilli I, Bacchetti T, Tiano L. Effect of ubiquinol supplementation on biochemical and oxidative stress indexes after intense exercise in young athletes. *Redox Rep*. 2018; 23(1):136-145.
- Park SY, Pekas EJ, Headid III RJ, Son WM, Wooden TK, Song J, Layec G, Yadav SK, Mishra PK, Pipinos II. Acute mitochondrial antioxidant intake improves endothelial function, antioxidant enzyme activity, and exercise tolerance in patients with peripheral artery disease. *Am J Physiol Heart Circ Physiol*. 2020; 319:456-467.
- Cho SY, So WY, Roh HT. Effect of C242T Polymorphism in the Gene Encoding the NAD(P)H Oxidase p22^{phox} Subunit and Aerobic Fitness Levels on Redox State Biomarkers and DNA Damage Responses to Exhaustive Exercise: A Randomized Trial. *Int J Environ Res Public Health*. 2020; 17(12):4215-4226.
- Yimcharoen M, Kittikunnathum S, Suknikorn C, Nak-on W, Yeethong P, Anthony TG, Bunpo P. Effects of ascorbic acid supplementation on oxidative stress markers in healthy women following a single bout of exercise. *J Int Soc Sports Nutr*. 2019; 16(1):1-9.
- Morrison D, Hughes J, della Gatta PA, Mason S, Lamon S, Russel AP, Wadley GD. Vitamin C and E supplementation prevents some of the cellular adaptations to endurance-training in humans. *Free Radic Biol Med*. 2015; 89:852-862.
- Vezzoli A, Dellanoce C, Mrakic-Sposta S, Montorsi M, Moretti S, Tonini A, Pratali L, Accinni R. Oxidative Stress Assessment in Response to Ultraendurance Exercise: Thiols Redox Status and ROS Production according to Duration of a Competitive Race. *Oxid Med Cell Longev*. 2016; 2016:1-13.

11. Koenig RT, Dickman JR, Kang CH, Zhang T, Chu YF, Ji LL. Avenanthramide supplementation attenuates eccentric exercise-inflicted blood inflammatory markers in women. *Eur J Appl Physiol*. 2016; 116:67-76.
12. Trewin, A. J., Lundell, L. S., Perry, B. D., Patil, K. V., Chibalin, A. V., Levinger, I., McQuade, L. R., Stepto, N. K. Effect of N-acetylcysteine infusion on exercise-induced modulation of insulin sensitivity and signaling pathways in human skeletal muscle. *Am J Physiol Endocrinol Metab*. 2015; 309(4):388-397.
13. Martin N, Smith AC, Dungey MR, Young HML, Burton JO, Bishop NC. Exercise during hemodialysis does not affect the phenotype or prothrombotic nature of microparticles but alters their proinflammatory function. *Physiol Rep*. 2018; 6(19):1-13.
14. Jenkins NT, Landers RQ, Thakkar SR, Fan X, Brown MD, Prior SJ, Spangenburg EE, Hagberg JM. Prior endurance exercise prevents postprandial lipaemia-induced increases in reactive oxygen species in circulating CD31 + cells. *J Physiol*. 2011; 589(22):5539-5553.
15. Petersen AC, McKenna MJ, Medved I, Murphy KT, Brown MJ, Gatta PD, Cameron-Smith D. Infusion with the antioxidant N-acetylcysteine attenuates early adaptive responses to exercise in human skeletal muscle. *Acta Physiologica*. 2012; 204(3):382-392.
16. Mrakic-Spota S, Gussoni M, Montorsi M, Porcelli S, Vezzoli A. Assessment of a Standardized ROS Production Profile in Humans by Electron Paramagnetic Resonance. *Oxid Med Cell Longev*. 2012; 2012:1-10.
17. Dal Negro RW, Visconti M. Erdosteine reduces the exercise-induced oxidative stress in patients with severe COPD: Results of a placebo-controlled trial. *Pulmonary Pharmacology & Therapeutics*. 2016; 41:48-51.
18. Place N, Ivarsson N, Venckunas T, Westerblad H. Ryanodine receptor fragmentation and sarcoplasmic reticulum Ca²⁺ leak after one session of high-intensity interval exercise. *PNAS*. 2015; 112(50):15492-15498.
19. Hajizadeh MB, Tartibian B, Chehrizi M. The effects of three different exercise modalities on markers of male reproduction in healthy subjects: a randomized controlled trial. *Reproduction*. 2017; 153(2):157-174.
20. Pileggi CA, Hedges CP, D'Souza RF, Durainayagam BR, Markworth JF, Hickey AJR, Mitchell CJ, Cameron-Smith D. Exercise recovery increases skeletal muscle H₂O₂ emission and mitochondrial respiratory capacity following two-weeks of limb immobilization. *Free Rad Biol and Med*. 2018; 124:241-248.
21. Clifford T, Bowman A, Capper T, Allerton DM, Foster E, Birch-Machin M, Lietz G, Howatson G, Stevenson EJ. A pilot study investigating reactive oxygen species production in capillary blood after a marathon and the influence of an antioxidant-rich beetroot juice. *Appl Physiol Nutr Metab*. 2018; 43(3):303-306.
22. Caruana H and Marshall JM. Effects of modest hyperoxia and oral vitamin C on exercise hyperaemia and reactive hyperaemia in healthy young men. *Eur J Appl Physiol*. 2015; 115(9):1995-2006.
23. Harms-Ringdahl M, Jenssen D, Haghdoost S. Tomato juice intake suppressed serum concentration of 8-oxodG after extensive physical activity. *Nutr J*. 2012; 11(29):1-5.
24. White SH and Warren LK. Submaximal exercise training, more than dietary selenium supplementation, improves antioxidant status and ameliorates exercise-induced oxidative damage to skeletal muscle in young equine athletes. *J Anim Sci*. 2020; 98(7):1-5.
25. Garten R, Goldfarb A, Crabb B, Waller J. The Impact of Partial Vascular Occlusion on Oxidative Stress Markers during Resistance Exercise. *Int J Sports Med*. 2015; 36(07):542-549.
26. Ranadive SM, Joyner MJ, Walker BG, Taylor JL, Casey DP. Effect of vitamin C on hyperoxia-induced vasoconstriction in exercising skeletal muscle. *J Appl Physiol*. 2014; 117(10):1207-1211.
27. Goulart MJVC, Pisamiglio DS, Moller GB, Dani C, Alves FD, Bock PM, Schneider CD. Effects of grape juice consumption on muscle fatigue and oxidative stress in judo athletes: a randomized clinical trial. *An Acad Bras de Ciênc*. 2020; 92(4):1-14.
28. Hajizadeh Maleki B, Tartibian B. High-intensity interval training modulates male factor infertility through anti-inflammatory and antioxidative mechanisms in infertile men: A randomized controlled trial. *Cytokine*. 2020; 125:154861.
29. Cobley JN, McGlory C, Morton JP, Close GL. N-Acetylcysteine's Attenuation of Fatigue After Repeated Bouts of Intermittent Exercise: Practical Implications for Tournament Situations. *Int J Sport Nutr Exerc Metab*. 2011; 21(6):451-461.
30. Dopheide, J. F., Scheer, M., Doppler, C., Obst, V., Stein, P., Vosseler, M., Abegunewardene, N., Gori, T., Münzel, T., Daiber, A., Radsak, M. P., Espinola-Klein, C. Change of walking distance in intermittent claudication: impact on inflammation, oxidative stress and mononuclear cells: a pilot study. *Clin Res in Cardiol*. 2015; 104(9):751-763.
31. Ahmad NS, Abdul Aziz A, Kong KW, Hamid MSA, Cheong JPG, Hamzah SH. Dose-Response Effect of Tualang Honey on Postprandial Antioxidant Activity and Oxidative Stress in Female Athletes: A Pilot Study. *J Altern Complementary Med*. 2017; 23(12):989-995.
32. Sanguigni V, Manco M, Sorge R, Gnessi L, Francomano D. Natural antioxidant ice cream acutely reduces oxidative stress and improves vascular function and physical performance in healthy individuals. *Nutrition*. 2017; 33:225-233.
33. Broome SC, Braakhuis AJ, Mitchell CJ, Merry TL. Mitochondria-targeted antioxidant supplementation improves 8 km time trial performance in middle-aged trained male cyclists. *J Int Soc Sports Nutr*. 2021; 18(1):1-11.
34. Vasconcelos Gouveia, S. S., Pertinni de Morais Gouveia, G., Souza, L. M., Cunha da Costa, B., Iles, B., Pinho, V. A., Vasconcelos, S. S., Rolim Medeiros, J. V., Lago da Silva, R., Porto Pinheiro, L. G. The effect of pilates on metabolic control and oxidative stress of diabetics type 2 – A randomized controlled clinical trial. *J Bodyw Mov Ther*. 2021; 27:60-66.
35. Tolahunase M, Sagar R, Dada R. Impact of Yoga and Meditation on Cellular Aging in Apparently Healthy Individuals: A Prospective, Open-Label Single-Arm Exploratory Study. *Oxid Med Cell Longev*. 2017; 2017:1-9.
36. Yee MM. A Comparative Studies on Antimicrobial Activity and Antioxidant Activity on Different Extracts of Leaf, Bark and Root of *Moringa oleifera* Lamk (Drumstick tree). *Int J of Recent Innov Acad Res*. 2019; 3(7):24-34.
37. Ezeamuzie IC, Ambakederemo AW, Shode FO, Ekwebelem SC. Antiinflammatory effects of *Moringa oleifera* root extract. *Pharm Biol*. 1996; 34(3):207-212.
38. Okereke CJ and Akaninwor JO. The protein quality of raw leaf, seed and root of *Moringa oleifera* grown in Rivers State, Nigeria. *Scholars Research Library Annals of Biological Research*. 2013; 4(11):34-38.
39. Faizi S, Siddiqui BS, Saleem R, Aftab K, Shaheen F, Gilani AH. Hypotensive constituents from the pods of *Moringa oleifera*. *Planta Med*. 1998; 64:225-228.
40. Paliwal R, Sharma V, Pracheta, Sadhna S. Elucidation of free radical scavenging and antioxidant activity of aqueous and hydro-ethanolic extract of *Moringa oleifera* pods. *Res J Pharm Technol*. 2011; 4(4):566-571.
41. Sharma V, Paliwal R, Janmeda P, Sharma S. Renoprotective effects of *Moringa oleifera* pods in 7, 12 dimethylbenz [a] anthracene-exposed mice. *J Chin Integr Med*. 2012; 10(10):1171-1178.
42. Paikra BK, Dhongade HKJ, Gidwani B. Phytochemistry and pharmacology of *Moringa oleifera* Lam. *J Pharmacopunct*. 2017; 20(3):194-200.
43. Kou X, Li B, Olayanju JB, Drake JM, Chen N. Nutraceutical or pharmacological potential of *Moringa oleifera* Lam. *Nutrients*. 2018; 10(3):1-12.
44. Bhattacharya A, Tiwari P, Sahu PK, Kumar S. A review of the phytochemical and pharmacological characteristics of *Moringa oleifera*. *J Pharm Bioallied Sci*. 2018;10(4):181-191.

45. Rani NZA, Husain K, Kumolosasi E. *Moringa* genus: A review of phytochemistry and pharmacology. *Front Pharmacol*. 2018; 9:1–26.
46. Saa RW, Fombang EN, Ndjantou EB, Njintang NY. Treatments and uses of *Moringa oleifera* seeds in human nutrition: A review. *Food Sci Nutr*. 2019; 7(6):1911–1919.
47. Baldisserotto A, Buso P, Radice M, Disette V, Lampronti I, Gambari R, et al. *Moringa oleifera* leaf extracts as multifunctional ingredients for “natural and organic” sunscreens and photoprotective preparations. *Molecules*. 2018; 23(3):1–16.
48. Ma ZF, Ahmad J, Zhang H, Khan I, Muhammad S. Evaluation of phytochemical and medicinal properties of *Moringa (Moringa oleifera)* as a potential functional food. *S Afr J Bot*. 2020; 129:40–46.
49. Bharali R, Tabassum J, Azad MRH. Chemomodulatory effect of *Moringa oleifera*, Lam, on hepatic carcinogen metabolizing enzymes, anti-oxidant parameters and skin papillomagenesis in mice. *Asian Pac J Cancer Prev*. 2003; 4(2):131–139.
50. Saini RK, Sivanesan I, Keum YS. Phytochemicals of *Moringa oleifera*: a review of their nutritional, therapeutic and industrial significance. *3 Biotech*. 2016; 6(2):1–14.
51. Rashid U, Anwar F, Moser BR, Knothe G. *Moringa oleifera* oil: A possible source of biodiesel. *Bioresour Technol*. 2008; 99(17): 8175–8179.
52. Karthivashan G, Arulselvan P, Wei-tan S, Fakurazi S. The molecular mechanism underlying the hepatoprotective potential of *Moringa oleifera* leaves extract against acetaminophen induced hepatotoxicity in mice. *J Funct Foods*. 2015; 17:115–126.
53. Meireles D, Gomes J, Lopes L, Hinzmann M, Machado J. A review of properties, nutritional and pharmaceutical applications of *Moringa oleifera*: integrative approach on conventional and traditional Asian medicine. *Adv Trad Med*. 2020; 20:495–515.
54. Fitriana WD, Ersam T, Shimizu K, Fatmawati S. Antioxidant activity of *Moringa oleifera* extracts. *Indones J Chem*. 2016; 16:297–301.
55. Nwidu LL, Elmorsy E, Aprioku JS, Siminialayi I, Carter WG. In Vitro Anti-Cholinesterase and Antioxidant Activity of Extracts of *Moringa oleifera* Plants from Rivers State, Niger Delta, Nigeria. *Medicines*. 2018; 5(3):1–17.
56. Karadi RV, Gadge NB, Alagawadi KR, Savadi RV. Effect of *Moringa oleifera* Lam. root-wood on ethylene glycol induced urolithiasis in rats. *J Ethnopharmacol*. 2006; 105:306–311.
57. Junior A, Sanon PJ, Lorde D. Phenotypic diversity of Haitian Benzolive (*Moringa oleifera* Lam.). *Pla Sci*. 2020; 03:1–6.
58. Cruz da Silva AV, Ferreira dos Santos AR, Da- Silva Lédo A, Feitosa RB, Almeida CS, Melo da Silva G, Rangel MSA. *Moringa* Genetic Diversity From Germplasm Bank Using RAPD Markers. *Trop Subtrop Agroecosystems*. 2012; 15:31–39.
59. Kleden MM, Soetanto H, Kusmartono K. Genetic diversity evaluation of *Moringa oleifera*, Lam from East Flores Regency using marker Random Amplified Polymorphic DNA (RAPD) and its relationship to chemical composition and in vitro gas production. *AGRIVITA J Agric Sci*. 2017; 39:219–231.
60. Raja S, Bagle BG, More TA. Drumstick (*Moringa oleifera* Lam.) improvement for semiarid and arid ecosystem: Analysis of environmental stability for yield. *J Plant Breed Crop Sci*. 2013; 5:164–170.
61. Mgendi MG, Nyomora AM, Manoko MK. Using morphological markers to assess variations between and within cultivated and non-cultivated provenances of *Moringa oleifera* Lam. in Tanzania. *J Life Sci*. 2011; 5:387–392.
62. Jaiswal, D., Rai, P. K., Mehta, S., Chatterji, S., Shukla, S., Rai, D. K., Sharma, G., Sharma, B., Khair, S., Watal, G. Role of *Moringa oleifera* in regulation of diabetes-induced oxidative stress. *Asian Pac J Trop Med*. 2013;6: 426–432.
63. Metwally FM, Rashad HM, Ahmed HH, Mahmoud AA, Raouf ERA, Abdalla AM. Molecular mechanisms of the anti-obesity potential effect of *Moringa oleifera* in the experimental model. *Asian Pac J Trop Biomed*. 2017; 7:214–221.
64. Babu PV, Liu D, Gilbert ER. Recent advances in understanding the anti-diabetic actions of dietary flavonoids. *J Nutr Biochem*. 2013; 24:1777–1789.
65. Oh YS and Jun HS. Role of bioactive food components in diabetes prevention: Effects on Beta-cell function and preservation. *Nutr Metab Insights*. 2014; 7:51–59.
66. Bahadoran Z, Mirmiran P, Azizi F. Dietary polyphenols as potential nutraceuticals in management of diabetes: A review. *J Diabetes Metab Disord*. 2013; 12:1–9.
67. Waterman C, Rojas-Silva P, Tumer TB, Kuhn P, Richard AJ, Wicks S, Stephens JM, Wang Z, Mynatt R, Cefalu W, Raskin I. Isothiocyanate-rich *Moringa oleifera* extract reduces weight gain, insulin resistance and hepatic gluconeogenesis in mice. *Mol Nutr Food Res*. 2015; 59:1013–1024.
68. Kim YJ and Kim HS. Screening *Moringa* species focused on development of locally available sustainable nutritional supplements. *Nutr Res Pract*. 2019; 13(6):529–534.
69. Cho AS, Jeon SM, Kim MJ, Yeo J, Seo KI, Choi MS, Lee MK. Chlorogenic acid exhibits anti-obesity property and improves lipid metabolism in high-fat diet-induced-obese mice. *Food Chem Toxicol*. 2010; 48:937–943.
70. Chen GL, Xu YB, Wu JL, Li N, Guo MQ. Hypoglycemic and hypolipidemic effects of *Moringa oleifera* leaves and their functional chemical constituents. *Food Chem*. 2020; 333:127478.
71. Lee YJ, Choi HS, Seo MJ, Jeon HJ, Kim KJ, Lee BY. Kaempferol suppresses lipid accumulation by inhibiting early adipogenesis in 3T3-L1 cells and zebrafish. *Food Funct*. 2015; 6:2824–2833.
72. Sangkitikomol W, Rocejanasaroj A, Tencomnao T. Effect of *Moringa oleifera* on advanced glycation end-product formation and lipid metabolism gene expression in HepG2 cells. *Genet Mol Res*. 2014; 13:723–735.
73. Sorrentino G, Ruggeri N, Specchia V, Cordenonsi M, Mano M, Dupont S, Manfrin A, Ingallina E, Sommaggio R, Piazza S, Rosato A, Piccolo S, Sal GD. Metabolic control of YAP and TAZ by the mevalonate pathway. *Nat Cell Biol*. 2014; 16:357–366.
74. Boopathy GTK and Hong W. Role of Hippo Pathway-YAP/TAZ Signaling in Angiogenesis. *Front Cell Dev Biol*. 2019; 7:1–12.
75. An Y, Kang Q, Zhao Y, Hu X, Li N. Lats2 Modulates Adipocyte Proliferation and Differentiation via Hippo Signaling. *PLoS One*. 2013; 8:1–10.
76. Piccolo S, Dupont S, Cordenonsi M. The Biology of YAP/TAZ: Hippo Signaling and Beyond. *Physiol Rev*. 2014; 94:1287–1312.
77. Panciera T, Azzolin L, Cordenonsi M, Piccolo S. Mechanobiology of YAP and TAZ in physiology and disease. *Nat Rev Mol Cell Biol*. 2017; 18:758–770.
78. Rausch V, Hansen CG. Review: The Hippo Pathway, YAP/TAZ, and the Plasma Membrane. *Trends Cell Biol*. 2020; 30:32–48.
79. Barbagallo I, Vanella L, Distefano A, Nicolosi D, Maravigna A, Lazzarino G, Di Rosa M, Tibullo D, Acquaviva R, Li Volti G. *Moringa oleifera* Lam. Improves lipid metabolism during adipogenic differentiation of human stem cells. *Eur Rev Med Pharmacol Sci*. 2016; 20:5223–5232.
80. Hwang JH, Kim AR, Kim KM, Park JI, Oh HT, Moon SA, Byun MR, Jeong H, Kim HK, Yaffe MB, Hwang ES, Hong J-H. TAZ couples Hippo/Wnt signalling and insulin sensitivity through Irs1 expression. *Nat Commun*. 2019; 10(1):1–11.
81. Tharp KM, Kang MS, Timblin GA, Dempersmier J, Dempsey GE, Zushin P-JH, Benavides J, Choi C, Li CX, Jha AK, Kajimura S, Healy KE, Sul HS, Saijo K, Kumar S, Stahl A. Actomyosin-Mediated Tension Orchestrates Uncoupled Respiration in Adipose Tissues. *Cell Metab*. 2018; 27:602–615.
82. Yamaguchi H and Taouk GM. A Potential Role of YAP/TAZ in the Interplay Between Metastasis and Metabolic Alterations. *Front Oncol*. 2020; 10:1–16.
83. Zheng Y and Pan D. The Hippo Signaling Pathway in Development and Disease. *Dev Cell*. 2019; 50:265–282.

84. Toma A and Deyno S. Phytochemistry and pharmacological activities of *Moringa oleifera*. Int J Pharm Sci Res. 2013; 1:222–231.
85. Tshabalala T, Ndhala AR, Ncube B, Abdelgadir HA, Van Staden J. Potential substitution of the root with the leaf in the use of *Moringa oleifera* for antimicrobial, antidiabetic and antioxidant properties. S Afr J Bot. 2020; 129:106–112.
86. Atta AH, Mounair SM, Nasr SM, Sedky D, Mohamed AM, Atta SA. Phytochemical studies and anti-ulcerative colitis effect of *Moringa oleifera* seeds and Egyptian propolis methanol extracts in a rat model. Asian Pac J Trop Biomed. 2019; 9:98–108.
87. Cretella ABM, Soley BS, Pawloski PL, Ruziska RM, Scharf DR, Ascari J, et al. Expanding the anti-inflammatory potential of *Moringa oleifera*: topical effect of seed oil on skin in inflammation and hyperproliferation. J Ethnopharmacol. 2020; 254:112708.
88. Vinoth B, Manivasagaperumal R, Balamurugan S. Phytochemical analysis and antibacterial activity of *Moringa oleifera*. Int J Med Pharm Sci. 2014; 4:27–34.
89. Sikder K, Das N, Kesh SB, Dey S. Quercetin and β - sitosterol prevent high fat diet induced dyslipidemia and hepatotoxicity in Swiss albino mice. Indian J Exp Biol. 2014; 52:60–66.
90. Ohsumi Y. Historical landmarks of autophagy research. Cell Res. 2014; 24:9–23.
91. Liao PC, Lai MH, Hsu KP, Kuo YH, Chen J, Tsai MC, Li CX, Yin XJ, Jeyashoke N, Chao LKP. Identification of β -Sitosterol as in Vitro Anti-Inflammatory Constituent in *Moringa oleifera*. J Agric Food Chem. 2018; 66:10748–10759
92. Ogbunugafor H, Igwo-Ezikpe M, Igwilo I, Ozumba N, Adenekan S, Ugochukwu C, Onyekwelu O, Ekechi A. *In vitro* and *In vivo* Evaluation of Antioxidant Properties of *Moringa Oleifera* Ethanolic Leaves Extract and Effect on Serum Lipid Indices in Rat. Maced J Med Sci. 2012; 5:397–403.
93. Fidrianny I, Kanapa I, Singgih M. Phytochemistry and Pharmacology of *Moringa* Tree: An Overview. Biointerface Res Appl Chem. 2021; 11:10776–10789.
94. Shanmugavel G, Prabakaran K, George B. Evaluation of phytochemical constituents of *Moringa oleifera* (Lam.) leaves collected from Puducherry region, South India. Int J Zool Appl Biosci. 2018; 3:1–8.
95. Hagoel L, Vexler A, Kalich-Philosoph L, Earon G, Ron I, Shtabsky A, Marmor S, Lev-Ari S. Combined effect of *Moringa oleifera* and ionizing radiation on survival and metastatic activity of pancreatic cancer cells. Integr Cancer Ther. 2019; 18:1–11.
96. Arif M, Yustisia I, Padliannah. The combination from ethanol extract of *Moringa* leaves (*Moringa oleifera* L.) and ethanol extract of papaya leaves (*Carica papaya* L.) slows the tumor growth in sprague dawley rats induced 7,12-dimethylbenz(a)anthracene. Med Cli Pract. 2020; 3:1–3.
97. Brown A, Emrani J, Mowa CN, Ahmed. *Moringa oleifera* and vesicular stomatitis virus : A combination approach for the treatment of cervical cancers. S Afr J Bot. 2020; 129:388–396.
98. Cuellar-Nuñez ML, Luzardo-Ocampo I, Campos-Vega R, Gallegos-Corona MA, González de Mejía E, Loarca-Piña G. Physicochemical and nutraceutical properties of moringa (*Moringa oleifera*) leaves and their effects in an *in vivo* AOM/DSS-induced colorectal carcinogenesis model. Food Res Int. 2018; 105:159–168.