Paper

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Submission date: 10-Jul-2021 09:05AM (UTC+0700)

Submission ID: 1617734819

File name: Int_Publish_2017-1_Gondokesumo_MCBS-Wahyu_FKUKM.pdf (1.11M)

Word count: 4313 Character count: 22551

RESEARCH ARTICLE



α -/ β -Glucosidase and α -Amylase Inhibitory Activities of Roselle (*Hibiscus sabdariffa* L.) Ethanol Extract

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Background: Diabetes mellitus is a metabolic disease, characterized by hyperglycemia due to disturbance in both insulin secretion and function. One of theurapeutic approaches is to reduce blood glucose levels by inhbiting α -/ β -glucosidase and α -amylase involved in carbohydrate digestion. Thus, inhibition of these enzymes play important role in the treatment of diabetes mellitus. Roselle (*Hibiscus sabdariffa* L.) has been known to have several medicinal properties and potency as an antidiabetics agents. This research aimed to observe antidiabetic properties of roselle ethanol extract (REE) towards α -glucosidase, β -glucosidase and α -amylase.

Methods: REE was done with maceration technique using diluent of 70% ethanol. Antidiabetic properties were measured by inhibitory activity of α -amylase, α -glucosidase and β -glucosidase.

Results: REE was able to inhibit α -/ β -glucosidase and α -amylase in the highest concentration with inhibition percentage of 72.68, 47.34 and 73.08% respectively, and were comparable with Acarbose of 81.49, 50.97, 73.08%. The median inhibitory concentration (IC₅₀) of α -/ β -glucosidase and α -amylase of REE were 15.81, 41.77, 18.09 µg/mL respectively, and Acarbose were 9.45, 22.57, 3.64 µg/mL respectively.

Conclusions: REE inhibits α -/ β -glucosidase and α -amylase.

Keywords: Roselle, Acarbose, α -glucosidase, β -glucosidase, α -amylase, antidiabetic

Introduction

Diabetes mellitus (DM) is a metabolic disease, characterized by hyperglycemia due to disturbance in both insulin secretion and function. Type 1 DM is a type with defective insulin secretion by pancreatic β -cells, and type 2 DM is a type with insulin resistance (a condition in which peripheral cells do not respond normally to insulin) or β -cell dysfunction. DM has postprandial phase indicated by increased blood glucose levels, which is called postprandial hyperglycemia.

Antihiperglycemia is commonly used to reduce hyperglycemia prostaglandins via inhibition of responsible enzymes in glucose absorption that hydrolize carbohydrates, such as α -/ β -glucosidase and α -amylase.⁴ Glucosidase is an enzyme involved in complex carbohydrate dissociation in the small intestine that play its role to cleave glicosidic bonds, resulting in glucose release from the non-reducing end of an oligo- or poly-saccharide chain involved in glycoprotein biosynthesis.⁴

Common drugs are expensive or generate adverse

Date of submission: May 3, 2016 Accepted for publication: January 10, 2017

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Gondokesumo MA. et al. Antidiabetic Roselle Ethanol Extract

effects. Common adverse effects of enzyme-inhibitory drugs, like Acarbose, are excessive inhibition of pancreatic α -amylase, which can result in abdominal distention, flatulence and diarrhea. Thus, natural medicine is proposed to be an alternative. Traditional herbal medicine is believed to possess hypoglycemic activity. There are more than 800 species of plants reported to contain hypoglycemic properties.

Roselle (Hibiscus sabdariffa L) has been used as a medicinal beverage in many countries such as Australia, India, Myanmar, Thailand, Senegal, France, Gambia, Nigeria, Greece, Saudi Arabia, Sudan, Latin America, Panama, Indonesia, Malaysia and China.7 Bioactive compounds is documented in roselle such as alkaloids, anthocyanins, flavonoids, saponins, steroids, sterols and tannins.8 Roselle extract demonstrate anti-insulin resistance properties due to its activity in reducing hyperglycemia and hyperinsulinemia, as well as decrease in Low Densisty Lipoprotein (LDL) and High Density Lipoprotein (HDL).9 Roselle extract also play role in lowering blood glucose levels in streptozotocin-induced dibetic mice, it is comparable with glibenclamide. 6,10,11 Therefore our current study was carried out to evaluate possible effectivity of Roselle extract to inhibit α/β -glucosidases and α -amylase.

Materials and methods

Preparation roselle extract

Extraction was performed based on maceration method. ^{12,13} Roselle calyx were collected from Cimenyan, West Java, Indonesia. The plant was identified by herbarium staff of Department of Biology, School of Life Sciences and Technology, Bandung Institute of Technology, Indonesia. Dried and milled roselle calyx were soaked in 70% distillated ethanol and filtered every 24 hours. The process was performed until colorless filtrate was gained, then filtrate was evaporated to obtain ethanol extract. Roselle ethanol extract (REE) was stored at -20°C.

3 α-glucosidase inhibitory activity assay

The α-glucosidase inhibitor activity was tested with the modified method. 1.14,15 Mixture reaction consists of 4-Nitrophenyl α-D-glucopyranoside (N1377, Sigma-Aldrich, St.Louis, USA), phospate buffer (pH 7.0) and 3 mple solution. After mixed well, α-glucosidase was added, incubated at 37°C for 30 minutes, then Na,CO₃ was added.

The 4-Nitrophenyl α -D-glucopyranoside was measured at 400 nm wavelength with microplate reader The inhibition

400 nm wavelength with microplate reader The inhibition percentage of α -glucosidase was calculated according to equation 1:

% inhibition = $\overline{\text{(C-S) x 100}}$

C : absorbance of control S : absorbance of sample

3 β-glucosidase inhibitory activity assay

The β-glucosidase inhibitory activity assay was conducted using modified method. Mixture solution contained 4-Nitrophenyl β-D-glucopyranoside (N7006, Sigma-Aldrich), phosphate but r (pH 7.0) and sample solution. Mixture reaction was incubated at 37°C for 5 minutes, added with enzyme mixture and incubated for 15 minutes. Reaction was stopped by addition of $\frac{1}{2}$ CO₃. Absorbance of 4-Nitrophenyl β-D-glucopyranoside was measured at 400 nm wavelength. The inhibition percentage of β-glucosidase was calculated according to equation 1.

2 α-amylase inhibitory activity assay

The α-amylase inhibitory activity assay was perfor 2 dusing a modified method. 16 Sample was introduced into sample well, dimethyl sulfoxide (DMSO) was used as a blank. Fighermore, α-amylase enzyme (A7595, Sigma-Aldrich) was added into each well, except for blank well. Then the mixture was incubated at 37°C for 10 minutes, and added with starch solution in each well, while control 2 ell was added with phospate buffer. Another incubation at 37°C for 15 minutes was conducted. Enzymatic reaction was stopped by adding acidic iodine solution in each well. The absorbance was measured at 565 nm wavelength. The inhibition percentage of α-amylase was calculated according to equation 1.

2 Statistical analysis

Data was presented as mean \pm standard deviation. To compare treatments, analysis of variance (ANOVA) was used, and p<0.05 were considered as statistically significant, along with Duncan Post-Hoc Test significant and 95% confidence interval. The median inhibitory concentration (IC₅₀) was measured to determine the inhibitory activies of α/β -glucosidase, α -amylase, according to linear regression. SPSS version 20.0 program was used for statistical analysis.

Results

a-glucosidase inhibitory activity

The α -glucosidase inhibitory of REE and Acarbose can be seen in Table 1 and Figure 1, and while the value of IC $_{50}$ can be seen in Table 2.

α-glucosidase activity of REE and Acarbose occurred in concentration-dependent manner in which higher α-glucosidase inhibition present in higher concentration of sample. Figure 1 shows that at the highest concentration (37.5 μg/mL), Acarbose generated higher inhibitory activity, compared with REE. The IC₅₀ value of Acarbose 14.65 μM or 9.45 μg/mL was more active than REE 15.81 μg/mL (Table 2). Inhibition of α-glucosidase is expected to normalize blood glucose level. ¹⁷ Based on statistical analysis, both REE and Acarbose had significant α-glucosidase inhibition activity differences at each concentration, higher concentration increased the activity (p<0.05).

5 β-glucosidase inhibitory activity

The β -glucosidate is a carbohydrate hydrolizing enzyme associated with metabolic disorder such as DM. Inhibition of carbohydrate hydrolyzing enzymes is used as therapeutic approach to decrease hyperglucose level and improve hyperglicemic. The β -glucosidase inhibitory of REE and Acarbose can be seen in Table 3 and Figure 2, the IC value can be seen in Table 4. The result of β -glucosidase activity in REE and Acarbose showed that inhibitory

Table 1. Effect concentrations toward α-glucosidase inhibitory activity (%) of REE and Acarbose

| Inhibitory Activity (%) | | |
|-------------------------|--|--|
| REE | Acarbose | |
| 72.68±1.56 ^e | 81.49±1.19 ° | |
| 46.38 ± 2.72^{d} | 44.94±0.54 | |
| 41.63±0.74 ° | 38.21±0.32 ° | |
| 38.79±0.26 b | 34.86±0.43 a | |
| $33.16{\pm}1.00~^{a}$ | 32.82±0.83 ° | |
| 32.48±1.05 a | 32.44±2.82 a | |
| | 72.68±1.56 ° 46.38±2.72 d 41.63±0.74 ° 38.79±0.26 b 33.16±1.00 a | |

The data are presented as mean±standard deviation. Different superscript small letters (*he.d.e*) indicate significance in the same column among concentrations of samples, used Duncan's post hoc test at p<0.05. Each treatment of samples was performed in triplicate.

activity was associated with concentrations in which higher β -glucosidase inhibition present in higher concentration of sample. Inhibition percentage of Acarbose with percentage 50.97 ± 3.78 was comparable with REE 47.34±2.69 (Table 3). As shown in Table 4, Acarbose was more active in inhibiting β -glucosidase, the IC $_{50}$ value 34.96 μM or 22.57 $\mu g/mL$ was lower compared to REE 41.77 $\mu g/mL$. Statistical analysis showed REE and Acarbose had significant differences in β -glucosidases inhibition activity at each concentration, higher concentration increased β -glucosidase inhibition ($p{<}0.05$).

a-amylase inhibitory activity

Pancreatic α-amylase is an enzyme in the digestive system and catalyses the initial step in starch hydrolysis. ¹⁹ Inhibition of intestinal α-amylase delays the starch and oligosaccharides degradation to monosaccharides prior to absorbtion, which later decreases the glucose absorption, as well as postprandial blood glucose level. ^{20,21} The α-amylase activity showed inhibitory activity in concentration-dependent manner in which increasing concentrations generated higher inhibitory activity. Inhibitory activity was lower in REE compared to Acarbose (Table 5 & 6, Figure 3) with inhibition percentage 73.08±3.85 of REE while Acarbose 77.08±4.77. Acarbose was more active in inhibiting α-amylase, the IC₅₀ value 5.64 μM or 3.64 μg/mL was more active compared to REE 18.09 μg/mL. Based on statistical analysis, both REE and Acarbose had significant

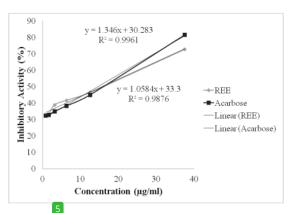


Figure 1. The α -glucosidase inhibitory activity (%) of REE and Acarbose. REE and Acarbose were diluted in 10% DMSO to reach the final concentrations of 37.5, 12.5, 6.25, 3.125, 1.563, 0.398, 0.781 μ g/mL.

Table 2. The median inhibitory activity (IC_{ea}) of α-glucosidase of REE and Acarbose

| Commis | Ecuation | R ² | IC | Assessed IC |
|-----------------|----------------------|----------------|--------------------------------|--|
| Sample | Equation | К | IC ₅₀ (μg/mL/μM) | Average IC ₅₀ ₍ μg/mL/μM) |
| REE Test 1 | Y = 1.0166x + 33.096 | 0.9866 | 16.63 μg/mL | |
| REE Test 2 | Y = 1.0296x + 33.828 | 0.9845 | 15.71 μg/mL | |
| REE Test 3 | Y = 1.1288x + 32.976 | 0.9802 | $15.08~\mu\text{g/mL}$ | |
| Average | y = 1.0584x + 33.3 | 0.9876 | 15.77 μg/mL | 15.81 ± 0.78 μg/mL |
| Acarbose Test 1 | Y = 1.3975x + 29.574 | 0.9977 | 14.62 μM | |
| | | | $(9.43~\mu g/mL)$ | |
| Acarbose Test 2 | Y = 1.3267x + 31.097 | 0.9870 | 14.25 μΜ | |
| | | | $(9.43~\mu g/mL)$ | |
| Acarbose Test 3 | Y = 1.3139x + 30.179 | 0.9955 | 15.09 μΜ | |
| | | | $(9.74~\mu g/mL)$ | |
| Average | y = 1.346X + 30.283 | 0.9961 | 14.64 μΜ | $14.65 \pm 0.42 \mu\text{M}$ |
| 3 | | | $(9.45~\mu g/mL)$ | (9,45 μg/mL) |

The $\overline{\text{IC}}_{s_0}$ value was presented as mean \pm standard deviation in each replication of samples (test 1, 2, 3) and average from 3 replication. The IC_{s_0} value for REE in $\mu g/\text{mL}$ meanwhile Acarbose both in μM and $\mu g/\text{mL}$.

Table 3. Effect concentrations toward β -glucosidase inhibitory activity (%) of REE and Acarbose

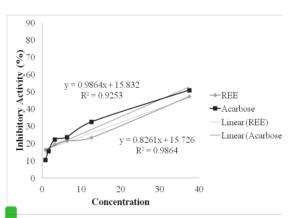
| Concentrations | Inhibitory Activity | | |
|-----------------|------------------------|--------------|--|
| (μg/mL or μM) — | REE | Acarbose | |
| 37.5 | 47.34±2.69 d | 50.97±3.78 ° | |
| 12.5 | 23.47±1.49 ° | 32.54±3.17 | |
| 6.25 | 21.66±0.62 bc | 23.81±0.23 | |
| 3.125 | $19.56{\pm}0.99~^{ab}$ | 22.35±1.06 ° | |
| 1.563 | 16.92±3.32 a | 15.60±1.00 1 | |
| 0.781 | 16.39±0.85 a | 10.60±1.93 ° | |

The data are presented as mean±standard deviation. Different superscript small letters (*a,b,b,b,c,c,d,e*) indicate significance in the same column among concentrations of samples, used Duncan's post hoc test at p<0.05. Each treatment of samples was performed in triplicate.

difference in α -amylases inhibition activity at each concentration, higher concentration increased α -amylase inhibition (p<0.05).

Discussion

The result of present study shaped that REE possessed antidiabetic activity, based on inhibitory activity of α -/ β -glucosidase and α -amilase. These results indicate the antidiabetic properties of REE was lower than Acarbose.



5 gure 2. The β-glucosidase inhibitory activity (%) of REE and Acarbose. REE and Acarbose were diluted in 10% DMSO to reach the final concentrations of 37.5, 12.5, 6.25, 3.125, 1.563, 0.398, 0.781 μg/mL.

The α -/ β -glucosidase are carbohydrate hydrolyzing enzymes that take place in metabolic disorder such as DM. Glucosidase cleaves the glycosidic bond that release glucose from the non-reducing end of an oligo- or poly-saccharide chain involved in glycoprotein biosynthesis. ²² Inhibition of carbohydrate hydrolyzing enzymes is therefore used as therapeutic approach to decrease hyperglycemia. ²³ This data is in accordance with previous study that the aqueous extract of roselle has the inhibitory activity of α -glucosidase as well

Table 4. The median inhibitory activity (IC_{so}) of β-glucosidase of REE and Acarbose

| Table 4. The median inhibitory activity (1C ₅₀) or p-glucosidase of REE and Acarbose | | | | |
|--|---|---|---|--|
| Equation | R ² | IC ₅₀ (μg/mL/μM) | Average IC ₅₀ (μg/mL/μM) | |
| Y = 0.9265x + 14.298 | 0.9722 | 38.53 μg/mL | | |
| Y = 0.8285x + 15.843 | 0.9652 | 41.23 μg/mL | | |
| Y = 0.7235x + 17.036 | 0.9944 | 45.56 μg/mL | | |
| y = 0.8264x + 15.726 | 0.9864 | 41.48 μg/mL | 41.77 ± 3.55 μg/mL | |
| Y = 1.0059x + 16.285 | 0.9240 | 33.52 μM | | |
| | | $(21.64~\mu\text{g/mL})$ | | |
| Y = 1.0905x + 15.473 | 0.9075 | 31.66 µM | | |
| | | (20.43 µg/mL) | | |
| Y = 0.8627x + 15.738 | 0.9381 | 39.71 μΜ | | |
| | | (25.63 µg/mL) | | |
| y = 0.9864x + 15.832 | 0.9253 | 34.63 μM (22.35 μg/mL) | 34.96 ± 4.22 μM (22.57 μg/mL) | |
| | Equation $Y = 0.9265x + 14.298$ $Y = 0.8285x + 15.843$ $Y = 0.7235x + 17.036$ $y = 0.8264x + 15.726$ $Y = 1.0059x + 16.285$ $Y = 1.0905x + 15.473$ $Y = 0.8627x + 15.738$ | Equation \mathbb{R}^2 $Y = 0.9265x + 14.298$ 0.9722 $Y = 0.8285x + 15.843$ 0.9652 $Y = 0.7235x + 17.036$ 0.9944 $y = 0.8264x + 15.726$ 0.9864 $Y = 1.0059x + 16.285$ 0.9240 $Y = 1.0905x + 15.473$ 0.9075 $Y = 0.8627x + 15.738$ 0.9381 | Equation R^2 IC_{50} (μg/mL/μM) Y = 0.9265x + 14.298 0.9722 38.53 μg/mL Y = 0.8285x + 15.843 0.9652 41.23 μg/mL Y = 0.7235x + 17.036 0.9944 45.56 μg/mL y = 0.8264x + 15.726 0.9864 41.48 μg/mL Y = 1.0059x + 16.285 0.9240 33.52 μM (21.64 μg/mL) (20.43 μg/mL) Y = 0.8627x + 15.738 0.9381 39.71 μM (25.63 μg/mL) (25.63 μg/mL) | |

The IC_{s_0} value was presented as mean \pm standard deviation in each replication of samples (test 1, 2, 3) and aver 3 e from 3 replication. The IC_{s_0} value for REE in $\mu g/mL$ meanwhile Acarbose both in μM and $\mu g/mL$.

Table 5. Effect concentrations toward a-amylase inhibitory activity (%) of REE and Acarbose

| Concentrations | Inhibitory Activity | | |
|-----------------|-------------------------|-------------------------|--|
| (μg/mL or μM) _ | REE | Acarbose | |
| 34.29 | 73.08±3.85 ° | 77.08±4.77 ^d | |
| 11.43 | $41.88\pm6.45^{\ b}$ | 58.82±5.88 ° | |
| 5.71 | $38.00\pm4.00^{\ b}$ | 55.83±3.82 ° | |
| 2.86 | 25.49±3.92 a | 52.67±4.16 ° | |
| 1.43 | 22.42±2.78 ^a | 43.86±1.75 b | |
| 0.71 | 20.24±2.73 a | 35.45±0.92 a | |

The data are presented as mean±standard deviation. Different superscript small letters (*\frac{a,b,c,d}{}\) indicate significance in the same column among concentrations of samples, used Duncan's post hoc test at *p*<0.05. Each treatment of samples was performed in triplicate.

as lowering blood glucose levels in DM animal models.²⁴ Roselle ethanol extract reduced the blood glucose levels on diabetic mice was comparable with n-hexane and ethyl acetate extract of roselle.⁶ The ability might be due to presence of dietary phytochemicals in roselle such as anthocyanins, flavonoids and phenolic acids.²⁵ Roselle also has variety content bioflavonoid such as prunin, myrciacitrin, 6-hydroxyl-flavonoids, 6-hydroxyluteolin, apigenin, and luteolin, that can lower blood glucose, and have inhibitory

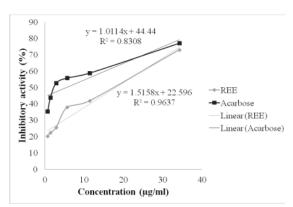


Figure 3. The α -amylase inhibitory activity (%) of REE and Acarbose. REE and Acarbose were diluted in 10% DMSO to reach the final concentrations of 37.5, 12.5, 6.25, 3.125, 1.563, 0.398, 0.781 μ g/mL.

activity of α -glucosidase.²⁴ Flavonoid contents of REE are flavonols (gossypetin) and the anthocyanins.²⁵ Flavonoid and isoflavonoid glycosides are found in human small intestine, in which α - and β -glucosidase inhibitory present to inhibit carbohydrate catalyses²⁶ that suggests contribution of flavonoid content as antidiabetic effects.

In this study, REE has α -amylase inhibitory activity. Inhibition of α -amylase will promote retardation of starch hydrolysis, that reduces digestion and absorption rate

Table 6. The median inhibitory activity (IC_{so}) of α -amylase of REE and Acarbose

| Sample | Equation | R ² | IC ₅₀ (μg/mL/μM) | Average IC ₅₀ (μg/mL/μM) |
|-----------------|----------------------|----------------|--------------------------------|--|
| REE Test 1 | Y = 1.5748x + 22.019 | 0.9858 | 17.77 μg/mL | |
| REE Test 2 | Y = 1.4414x + 23.607 | 0.8631 | 18.31 μg/mL | |
| REE Test 3 | Y = 1.5512x + 22.161 | 0.9454 | 18.18 μg/mL | |
| Average | Y = 1.5158x + 22.596 | 0.9637 | 18.07 μg/mL | $18.09 \pm~0.28~\mu\text{g/mL}$ |
| Acarbose Test 1 | Y = 0.8853x + 42.773 | 0.8291 | 8.16 μΜ | |
| | | | $(5.26~\mu g/mL)$ | |
| Acarbose Test 2 | Y = 1.0349x + 46.243 | 0.7196 | $3.63~\mu M$ | |
| | | | $2.10~\mu\text{g/mL}$ | |
| Acarbose Test 3 | Y = 1.1141x + 44.305 | 0.8909 | 5.11 μΜ | |
| | | | $(3.29~\mu\text{g/mL})$ | |
| Average | Y = 1.0114x + 44.44 | 0.8308 | 5.49 μM (3.54 μg/mL) | $5.64 \pm 2.31 \mu\text{M}$ (3.64 $\mu\text{g/mL}$) |

The IC_{s_0} value was presented as mean \pm standard deviation in each replication of samples (test 1, 2, 3) and average from 3 replication. The IC_{s_0} value for REE in $\mu g/mL$ meanwhile Acarbose both in μM and $\mu g/mL$.

of carbohydrates, resulting in decreased post-prandial hyperglycemia. 15 Flavanoids content in roselle play key role in α -amylase inhibitory activity. 27 Inhibition of α -amylase preserve β -cell integrity and function by removing free radicals could enhance protection against the progression of insulin resistance in type 2 DM. 28 Previous studies also showed that roselle extract has potency to inhibit α -amylase activity, sugars and starch absorption, which may assist in weight loss. 29

Flavonoids present in roselle, are well known to contain medicinal properties such as antiinflammatory, antioxidant and antidiabetic. 30,31 Phenolic and pronoids content in roselle are suggested to play role in inhibitory activity of both α -amylase and α - $/\beta$ -glucosidase. 32,33 Interactions between chemical compound of REE and enzymes causes alteration in the enzyme's molecular configuration, as well as hydrophilic and hydrophobic properties, that will decrease enzyme activities. 10 Active components of REE compete with the substrate in binding to active site of enzyme prevent the oligosaccharides breakdown to disaccharides. 34

Conclusion

REE is a promising antidiabetic agent to inhibit α -/ β -glucosidase and α -amylase. Further researches regarding

identification and development of bioactive compounds contained in roselle, are encouraged.

Conflicts of interest

All contributing authors declare no conflicts of interest.

Acknowledgements

The authors gratefully acknowledge the financial support from Research and Community Service of Surabaya University. This research was also supported by Biomolecular and Biomedical Research Center, Bandung, Indonesia. We are also thankful to Merry Afni, Ervi Afifah, Hayatun Nufus, Seila Arumwardana, Dwi Davidson Rahibiba for their valuable assistance.

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